

## Prevalence of *Enterocytozoon bieneusi* and *Blastocystis* sp. infection in foxes (*Alopex lagopus*) in northern China

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**Abstract** – To examine the prevalence of *E. bieneusi* and *Blastocystis* sp. in foxes in China, this study analyzed the prevalence and distribution of genotypes or subtypes. A total of 352 fresh fecal samples were collected from foxes across five provinces in northern China and analyzed using PCR. The overall prevalences of *E. bieneusi* and *Blastocystis* sp. were 48.3% and 2.0%, respectively; the highest prevalences were found in Shandong Province, with 87.1% and 5.4%, respectively. The prevalence rates were influenced by several factors; a breeding scale value <1,500 was related to higher prevalences. Multivariate analysis showed that the region and breeding scale were the main risk factors for *E. bieneusi*. Eleven genotypes of *E. bieneusi* were identified, all of which are classified within Group 1. This includes five previously characterized genotypes and six novel genotypes. Among these, CHN-F1 was the predominant genotype, accounting for 67.7% of cases. *Blastocystis* sp. was detected with only one subtype (ST3), which represents the first report of this genotype in foxes. The identification of *E. bieneusi* in foxes and the first detection of the ST3 subtypes of *Blastocystis* sp. contribute to a more comprehensive understanding of the epidemiology of these microeukaryotes. These findings suggest a potential pathway for the transmission of microeukaryotes from fox farms to human populations, underscoring the importance of monitoring the public health risks.

**Key words:** Microeukaryote, Genotypes, Subtypes, Zoonosis.

**Résumé** – Prévalence de l'infection par *Enterocytozoon bieneusi* et *Blastocystis* sp. chez les renards (*Alopex lagopus*) dans le nord de la Chine. Pour examiner la prévalence d'*E. bieneusi* et de *Blastocystis* sp. chez les renards en Chine, cette étude a analysé la prévalence et la distribution des génotypes ou sous-types. Au total, 352 échantillons de matières fécales fraîches ont été collectés auprès de renards dans cinq provinces du nord de la Chine et analysés par PCR. Les prévalences globales d'*E. bieneusi* et de *Blastocystis* sp. étaient respectivement de 48,3 % et de 2,0 %; les prévalences les plus élevées ont été trouvées dans la province du Shandong, avec respectivement 87,1 % et 5,4 %. Les taux de prévalence étaient influencés par plusieurs facteurs; une valeur d'échelle de reproduction < 1500 était liée à des prévalences plus élevées. Une analyse multivariée a montré que la région et l'échelle de reproduction étaient les principaux facteurs de risque pour *E. bieneusi*. Onze génotypes d'*E. bieneusi* ont été identifiés, tous classés dans le groupe 1. Ils incluent cinq génotypes précédemment caractérisés et six nouveaux génotypes. Parmi ceux-ci, le génotype CHN-F1 était prédominant, représentant 67,7 % des cas. *Blastocystis* sp. a été détecté avec un seul sous-type (ST3), ce qui représente le premier signalement de ce génotype chez les renards. L'identification d'*E. bieneusi* chez les renards et la première détection des sous-types ST3 de *Blastocystis* sp. contribuent à une compréhension plus complète de l'épidémiologie de ces microeucaryotes. Ces résultats suggèrent une voie potentielle de transmission des microeucaryotes des élevages de renards aux populations humaines, soulignant l'importance de surveiller les risques pour la santé publique.

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## Introduction

*Enterocytozoon bieneusi* and *Blastocystis* sp. are prevalent microeukaryotes [18, 33, 40, 67]. They can infect various hosts, including humans and foxes. Humans and animals become infected with *E. bieneusi* and *Blastocystis* sp. mainly through the fecal–oral ingestion of contaminated water or food [57]. Infections with two microeukaryotes are usually asymptomatic, but they can sometimes cause diarrhea [32, 67].

To date, at least 820 genotypes of *E. bieneusi* and 48 subtypes (STs) of *Blastocystis* sp. have been identified [32, 36, 39, 40, 54]. The *E. bieneusi* genotypes have been divided into 15 phylogenetic groups with different host specificity and zoonotic potential [32]. Group 1 is the largest, containing numerous zoonotic genotypes like D, Ebpc, and Peru 8, which are frequently detected in humans [24]. Even though certain genotypes like CHN-F1, NCF2, and NCF3 within Group 1 have not been detected in human patients thus far, they may still pose a zoonotic transmission risk [8, 32, 44]. The genotypes in Group 2 primarily infect ruminants, but also include the BEB4, BEB6, I, and J genotypes capable of infecting humans [36]. The remaining Groups 3–15 may exhibit host specificity, with some genotypes reported to have the ability to infect humans, such as Group 5 (KIN-3) and Group 6 (Nig 3 and May1) [15, 23]. More than one billion individuals worldwide are carriers of *Blastocystis* sp., with human infections involving ST1-10, ST12, ST14, ST16, ST23, ST35, and ST41 among the 48 identified subtypes. The prevalence of *Blastocystis* sp. infection in humans is predominantly attributed to ST1–4 which accounts for approximately 90.0% of cases [4, 21, 25, 29, 42, 65]. Although there are already reports on the prevalence of the *E. bieneusi* genotypes and the *Blastocystis* sp. subtypes, additional research is necessary to comprehensively understand their implications for public health [42].

A total of 361 *E. bieneusi* genotypes have been identified in China, including a variety of hosts, such as humans (5.7%, 188/3271, predominantly genotypes D and Ebpc), non-human primates (17.8%, 948/5318, predominantly genotype D), cattle (14.0%, 783/5594, predominantly genotypes J and I), sheep and goats (31.9%, 978/3068, predominantly genotype BEB6), pigs (55.9%, 1101/1969, predominantly genotypes Ebpc and EbpA), and blue foxes (17.9%, 108/603, predominantly genotype D) [53]. *Blastocystis* sp. is also widely distributed in China, the average infection rate among humans is 3.4% (3625/107,695), with ST3 being the predominant genotype [10]. Among other animals, the overall infection rate in Artiodactyla (including pigs, cattle, sheep, goats, reindeer, and sika and barking deer) is 27.4% (1050/3828), while in Carnivora (including raccoon domestic dogs and Arctic foxes), the infection rate is 2.8% (11/389) [10]. Another study involved a meta-analysis, showing that the global prevalence of *Blastocystis* sp. was 2.1% (29/1377) in free-living carnivores, compared to a significantly higher prevalence of 8.5% (100/1175) in captive individuals [5].

In China, the fox is an economically important animal, with approximately 3.44 million breeding foxes [66]. However, there is limited research on the prevalence of *E. bieneusi* and *Blastocystis* sp. in foxes, as well as their genetic characteristics. This study aimed to investigate the infection and distribution of

the *E. bieneusi* genotypes and the *Blastocystis* sp. subtypes in foxes in China. Understanding the genetic evolutionary relationship between *E. bieneusi* and *Blastocystis* sp. in foxes can help us to effectively prevent and control diseases among fox populations as well as safeguard public health in China.

## Materials and methods

### Ethics statement

All the procedures used in this study were approved by the Research Ethics Committee for the Care and Use of Laboratory Animals in Qingdao Agricultural University, China. Written informed consent was obtained from the owners of the animals involved in this study.

### Collection of samples

From October 2023 to June 2024, a total of 352 feces samples from foxes (*Alopex lagopus*) were collected across five provinces in northern China. Specifically, the samples were collected from three farms in Heilongjiang (sample number = 47, taken in 2024), two farms in Jilin (sample number = 40, taken in 2024), six farms in Liaoning (sample number = 10, taken in 2023; sample number = 69, taken in 2024), six farms in Hebei (sample number = 93, taken in 2024), and five farms in Shandong (sample number = 93, taken in 2023). Fresh fecal samples, representing approximately 1.0 to 1.5% of the total farmed population, were randomly collected from each farm. All the farms underwent biannual deworming treatments using either ivermectin or albendazole. Diarrhea is defined as the presence of green, watery, or bloody stools. The samples were collected from multiple locations within the farms to avoid being limited to a single area or cage. During the collection process, samples that met the defined criteria for diarrhea were recorded as diarrheal samples. The sampling process involved recording essential information, including the region, the breeding scale, the age of the fox, and the presence of diarrhea, which were documented during the data collection process. The collected samples were transported to a laboratory on ice within 24–48 h and stored at  $-80^{\circ}\text{C}$  until DNA extraction. The storage duration ranged from 1 week to 10 months.

### DNA extraction and PCR amplification

DNA extraction from all the samples was conducted utilizing an EZNAR Stool DNA Kit (OMEGA Biotek Inc., Norcross, GA, USA), with the material subsequently stored at  $-20^{\circ}\text{C}$ . The DNA of all the samples was amplified using PCR to determine the presence of *E. bieneusi*, or *Blastocystis* sp., based on the ITS region and the small subunit (SSU) rRNA gene [57, 59], respectively. Novoprotein 2×Taq Master Mix (Novoprotein Ltd., E005-01, Suzhou, China) was used for all the PCR amplifications. The reaction conditions and sizes of the product fragments are presented in Table 1. *Enterocytozoon bieneusi* (genotype D) and *Blastocystis* sp. (subtype 3)-positive DNA previously detected in a laboratory from wild rodents was used as the positive control, while dd H<sub>2</sub>O was employed as the

**Table 1.** Primers used in the characterization of *E. bienersi* and *Blastocystis* sp.

Gene	Primer (5'-3')	Product size (bp)	Annealing temperature (°C)
SSU rRNA of <i>Blastocystis</i> sp.	SSU rRNA-F: ATCTGGTTGATCCTGCCAGT	600	55
	SSU rRNA-R: GAGCTTTTAACTGCAACAACG		
ITS gene of <i>E. bienersi</i>	ITS-F1: GGTCATAGGGATGAAGAG	392	57
	ITS-R1: TTCGAGTTCTTTCGCGCTC		
	ITS-F2: GCTCTGAATATCTATGGCT		
	ITS-R2: ATCGCCGACGGATCCAAGTG		

negative control [15, 16]. The PCR products were visualized using the QuickGel 6200 gel imaging System (Mona Biotechnology Co., Ltd., GD50102, Suzhou, China) after electrophoresis on 1% agarose gels.

### Sequencing and phylogenetic analyses

All amplicons with the expected size were sequenced bidirectionally at the General Biol. Company in Anhui, China. Then, the obtained sequences were blasted in GenBank to finally confirm the positive samples (<http://www.ncbi.nlm.nih.gov/BLAST/>). The genotypes of *E. bienersi* were determined and named based on ~243 bp of the ITS region, according to the established nomenclature system [38]. Phylogenetic trees were constructed using the neighbor-joining (NJ) method in MEGA 11 (v. 11.0.13), with bootstrapping performed using 1,000 replicates to assess the genetic relationships among the *E. bienersi* genotypes. CD-HIT (Version 4.8.1) was employed to remove the redundant sequences, solely preserving the unique sequences at a 100% identity threshold. The unique sequences, which serve as representatives of their respective clusters, were subsequently submitted to GenBank (accession Nos. [PQ249072–PQ249088](#) and [PQ223428–PQ223432](#)).

### Statistical analysis

The chi-square test of SAS (SAS Institute Inc., Cary, NC, USA, v.9.4) was utilized to conduct statistical analysis of the region, the breeding scale, the age of the foxes, and the presence of diarrhea. Forward stepwise logistic regression, using the Fisher scoring technique, was applied to a binary Logit model to assess the impact of multivariate regression analysis on *E. bienersi* and *Blastocystis* sp. infection. All the tests were two-sided, and the results were considered statistically significant if  $p < 0.05$ , with corresponding odds ratios (ORs) and their 95% confidence intervals (95% CIs) provided [63].

## Results

### Prevalence of *Enterocytozoon bienersi* and *Blastocystis* sp.

This study identified a prevalence of 48.3% (170/352, 95% CI [42.97–53.65]) for *Enterocytozoon bienersi* and 2.0%

(7/352, 95% CI [0.80–4.05]) for *Blastocystis* sp. Among the 352 foxes examined (Tables 2 and 3). *Enterocytozoon bienersi* was detected across all five provinces, with the highest prevalence observed in Shandong (87.1%, 81/93, 95% CI [79.43–93.24]), followed by Liaoning (65.8%, 52/79, 95% CI [54.96–75.93]), Jilin (27.5%, 11/40, 95% CI [14.61–42.51]), Hebei (20.4%, 19/93, 95% CI [12.79–29.28]), and Heilongjiang Province (14.9%, 7/47, 95% CI [5.93–26.70]) (Table 2). The occurrence of *Blastocystis* sp. was relatively low, being detected solely in Shandong and Hebei Provinces, with infection rates of 5.4% (5/93, 95% CI [1.54–11.06]) and 2.2% (2/93, 95% CI [0.03–6.37]), respectively (Table 3).

Statistical analysis revealed that the infection rate of *E. bienersi* was significantly higher in foxes with a breeding scale value of less than 1500 (54.3%, 83/153, 95% CI [43.60–62.10]) compared to those with a breeding scale value greater than 1500 (43.7%, 87/199, 95% CI 36.88–50.68) ( $p < 0.0001$ ). Notably, the prevalence of *E. bienersi* was 60.8% in diarrheal foxes, which is significantly higher than that in non-diarrheal foxes (45.0%, 125/278, 95% CI [39.15–50.85]) ( $p = 0.0156$ ). Additionally, the prevalence in juvenile foxes (68.8%, 117/170, 95% CI [61.64–75.59]) was significantly higher than that in adult foxes (29.1%, 53/182, 95% CI [22.73–35.95]) ( $p < 0.001$ , Table 2).

Similarly, the infection rate of *Blastocystis* sp. was higher in foxes with a breeding scale value of less than 1500 (3.3%, 5/153, 95% CI [0.93–6.79]) compared to those with a breeding scale value greater than 1500 (1.0%, 2/199, 95% CI [0.02–3.01]) ( $p = 0.0927$ ). The prevalence of *Blastocystis* sp. was 4.1% (3/74, 95% CI [0.51–10.02]) in diarrheal foxes, which is higher than in non-diarrheal foxes (1.4%, 4/278, 95% CI [0.31–3.25]) ( $p = 0.1673$ ). Furthermore, the prevalence in juvenile foxes (2.9%, 5/170, 95% CI [0.83–6.12]) was significantly higher than that in adult foxes (1.1%, 2/182, 95% CI [0.02–3.28]) ( $p = 0.2323$ , Table 3).

### Risk factors of *E. bienersi* and *Blastocystis* sp.

The region and breeding scale were significant in the final model, indicating its strong influence on *E. bienersi*. The region and breeding scale of foxes had a strong effect on *E. bienersi* infection in this study. The foxes in Shandong Province (OR = 0.408, 95% CI [0.339–0.491]) appear to exhibit a higher susceptibility to *E. bienersi* compared to those in Liaoning, Heilongjiang, Jilin, and Hebei Provinces. A breeding scale value  $< 1500$  (OR = 0.468, 95% CI [0.275–0.796]) indicates greater susceptibility to *E. bienersi* than a value  $\geq 1500$ . No

**Table 2.** Factors associated with prevalence of *E. bienersi* in foxes.

Factor	Category	No. positive/ tested	% (95% CI*)	Univariate meta-regression		Multivariate analysis <sup>c</sup>	
				OR (95% CI)	p-Value	OR (95% CI)	p-Value
Region	Heilongjiang Province	7/47	14.9 (5.93–26.70)	Reference <sup>d</sup>	<0.0001	0.408 (0.339–0.491)	<0.0001
	Jilin Province	11/40	27.5 (14.61–42.51)	2.17 (0.75–6.27)			
	Liaoning Province	52/79	65.8 (54.96–75.93)	11.05 (4.35–27.83)			
	Hebei Province	19/93	20.4 (12.79–29.28)	1.47 (0.57–3.79)			
	Shandong Province	81/93	87.1 (79.43–93.24)	39.52 (2.95–11.67)			
Breeding scale	<1500	83/153	54.3 (46.30–62.10)	1.53 (1.00–2.33)	0.0506	0.468 (0.275–0.796)	<0.0001
	≥1500	87/199	43.7 (36.88–50.68)	Reference			
Diarrhea	Yes	45/74	60.8 (49.39–71.68)	1.90 (1.13–3.21)	0.0156	–	–
	No	125/278	45.0 (39.15–50.85)	Reference			
Age of Fox	Adult <sup>a</sup>	53/182	29.1 (22.73–35.95)	Reference	<0.0001	–	–
	Juvenile <sup>b</sup>	117/170	68.8 (61.64–75.59)	12.38 (7.15–21.45)			
Total		170/352	48.3 (42.97–53.65)				

CI\*: confidence interval; a: ≥1 year old; b: <1 year old; c: Only effect factors are listed; d: control group for risk analysis.

**Table 3.** Factors associated with prevalence and subtypes of *Blastocystis* sp. in foxes.

Factor	Category	No. positive/tested	% (95% CI*)	Univariate meta-regression	
				p-Value	OR (95% CI)
Region	Heilongjiang Province	0/47	0 (0.00–3.63)	0.0927	–
	Jilin Province	0/40	0 (0.00–4.25)		–
	Liaoning Province	0/79	0 (0.00–2.16)		–
	Hebei Province	2/93	2.2 (0.03–6.37)		Reference <sup>c</sup>
	Shandong Province	5/93	5.4 (1.54–11.06)		2.59 (0.49–13.68)
Breeding scale	<1500	5/153	3.3 (0.93–6.79)	0.1439	3.33 (0.64–17.40)
	≥1500	2/199	1.0 (0.02–3.01)		Reference
Diarrhea	Yes	3/74	4.1 (0.51–10.02)	0.1673	Reference
	No	4/278	1.4 (0.31–3.25)		2.89 (0.63–13.23)
Age of Fox	Adult <sup>a</sup>	2/182	1.1 (0.02–3.28)	0.2323	Reference
	Juvenile <sup>b</sup>	5/170	2.9 (0.83–6.12)		2.73 (0.52–14.25)
Total		7/352	2.0 (0.80–4.05)		

CI\*: confidence interval; a: ≥1 year old; b: <1 year old; c: control group for risk analysis.

significant influencing factors were identified in the presence of *Blastocystis* sp. in the foxes.

### Distribution of *E. bienersi* genotypes and *Blastocystis* sp. subtypes

This study identified eleven genotypes of *E. bienersi* and one subtype of *Blastocystis* sp. among the 352 fox samples. The genotypes of *E. bienersi* comprise five known (D, Peru 8, WildBoar 3, CHN-F1, and CHN-DC1) and six newly identified genotypes (LNF-1, LNF-2, HBF-1, SDF-1, FJL-1, and FJL-2). These new genotypes contain a total of 11 polymorphic sites (Table 4). In this study, the recently discovered genotype CHN-F1 exhibited the highest prevalence rate, followed by NCF3, NCF2, and D. Type D was detected in Heilongjiang and Liaoning Provinces. Notably, CHN-F1 was detected in all the five provinces, whereas the remaining new genotypes LNF-1 and LNF-2 were found only in Liaoning Province, SDF-1 was detected in Shandong Province, and HBF-1 was found in Hebei Province. The ST3 subtype of *Blastocystis* sp. was exclusively detected in this study, limited to Hebei and Shandong Provinces (Fig. 1).

### Phylogenetic analysis

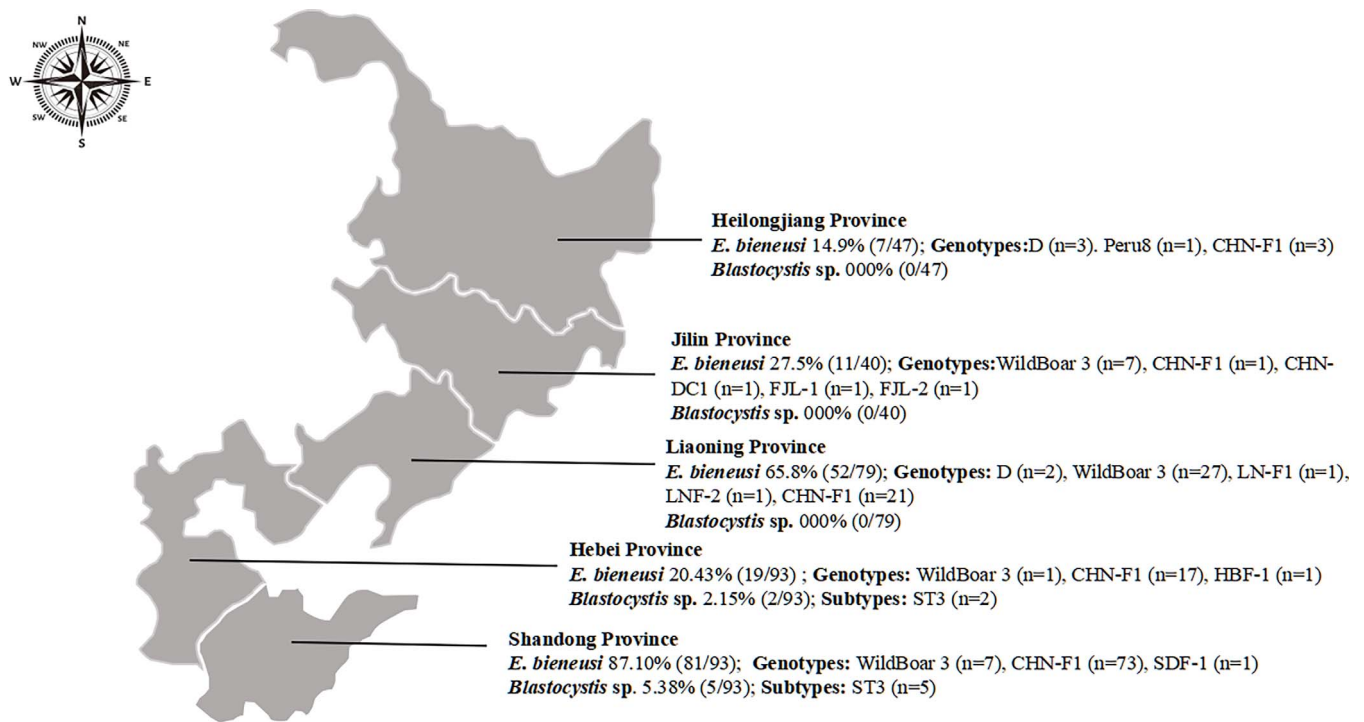
Phylogenetic trees were constructed to represent the eleven *E. bienersi* genotypes (Fig. 2) and the one *Blastocystis* sp. subtype identified in this study. It was observed that all the eleven *E. bienersi* genotypes were grouped in Group 1, with D, Peru8, and the novel genotypes LNF-1 and LNF-2 classified within Group 1a. The remaining detected genotypes CHN-F1, CHN-DC1, and WildBoar 3, as well as the newly discovered genotypes FJL-1, SDF-1, FJL-2, and HBF-1, were classified into Group 1b.

### Discussion

*Enterocytozoon bienersi* and *Blastocystis* sp. are significant zoonotic intestinal protozoa that have been understudied in foxes [1]. This study employed molecular epidemiologic methods to investigate the prevalence of these protozoa among foxes in northeastern China. Additionally, this study characterized the predominant *E. bienersi* genotypes and *Blastocystis* sp. subtypes in foxes and evaluated the potential zoonotic risk.

**Table 4.** Variations in the ~243 ITS nucleotide sequences among genotypes of the *E. bieneusi* in foxes.

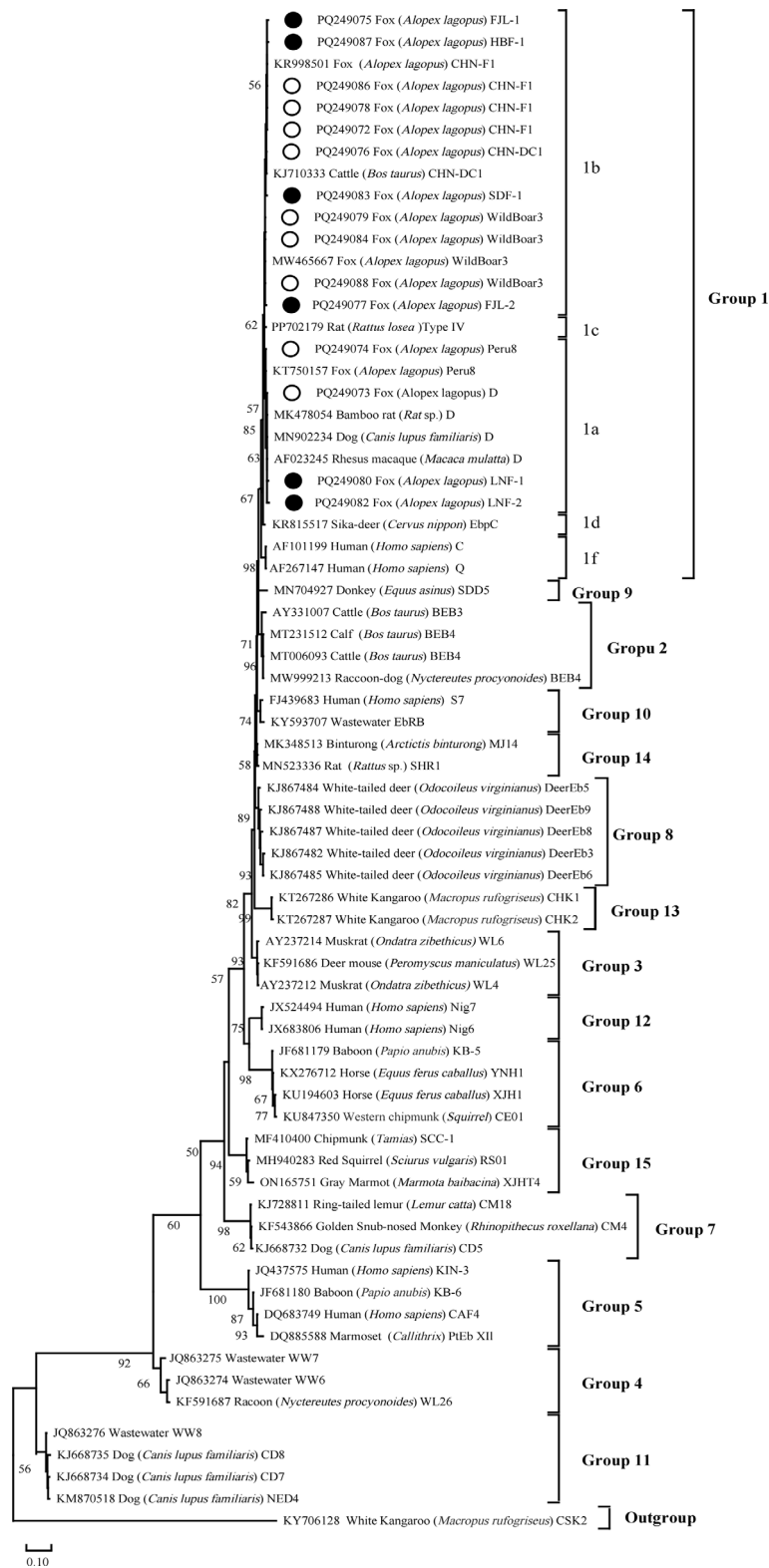
Genotypes (No.)	Nucleotide at position											GenBank accession No.
	17	44	48	65	77	93	117	131	191	209	212	
D (Reference)	G	T	T	G	G	C	T	G	G	T	G	AF023245 (Intergenic spacer)
D (5)												PQ249073
CHN-DC1 (1)				A		T	G	A				PQ249076
FJL-1 (1)	A	C		A		T	G	A				PQ249075
CHN-F1 (115)	A			A		T	G	A				PQ249072
HBF-1 (1)	A			A		T	G	A			A	PQ249087
LN-F1 (1)					A							PQ249080
LNF-2 (1)									A	C		PQ249082
FJL-2 (1)			C			T	G	A				PQ249077
WildBoar 3 (42)						T	G	A				PQ249079
Peru8 (1)							G					PQ249074
SDF-1 (1)	A						G	A				PQ249083



**Figure 1.** Distribution of *E. bieneusi* genotypes and *Blastocystis* sp. subtypes.

The prevalence of *E. bieneusi* was 48.3% (170/352), significantly higher than that reported in previous studies in foxes in China, such as 27.7% (53/191) [55], 16.4% (18/110) [66], and 12.3% (37/302) [62]. These studies employed the same PCR method as in the current study. In this study, the prevalence rate of *Blastocystis* sp. was 2.0%. Other studies have reported that the prevalence of *Blastocystis* sp. in foxes is 1.9% (4/213) in China [46] and 2.2% (4/213) in Spain [6]. These findings are consistent with the prevalence observed in our study. Strict carnivores tend to exhibit a lower prevalence of *Blastocystis* sp. than omnivorous or herbivorous species, and *Blastocystis* sp. in foxes may be caused by the contamination of food or water sources [5, 45].

Regional subgroup analysis revealed significant variation in *E. bieneusi* prevalence across the five provinces ( $p < 0.05$ ). The prevalence in this study is much higher than that in previous studies, such as in Jilin (7.7%) [62], Heilongjiang (7.1–27.7%) [55, 62], Hebei (17.7%) [62], and Xinjiang (2.9%) [64]. The previous studies reported 1.9% *Blastocystis* sp. prevalence in Jilin and Liaoning Provinces [46]. In comparison, the prevalence in Shandong and Hebei Provinces was higher than that given in previous reports. This study presents the first epidemiologic survey on *Blastocystis* sp. in foxes conducted in Hebei Province. Although it exhibited a low positive rate, this should still raise concerns among individuals. In provinces with a high prevalence of both *E. bieneusi* and *Blastocystis* sp., it is



**Figure 2.** Neighbor-joining tree of *E. bieneusi* ITS genotypes. Phylogenetic relationships of ITS nucleotide sequences of the *E. bieneusi* genotypes identified in this study and other reported genotypes. The phylogeny was inferred by a neighbor-joining analysis. Bootstrap values were obtained using 1000 replicates; those with values >50% are shown on the nodes. The genotypes in this study are marked by empty circles, and the novel genotypes are marked by filled circles.

important to monitor these microeukaryotes, although their veterinary significance and impact on foxes remain unclear. This includes ensuring strict adherence to environmental hygiene practices on farms and regular disinfection protocols, as well as routine testing. Furthermore, this study revealed a correlation between the breeding scale and the prevalence of these two protists, with farms having a breeding scale value <1500 exhibiting higher prevalence than those with a breeding scale value  $\geq 1500$ . This could be attributed to the fact that large-scale farms possess greater expertise in feeding management and disease prevention and control [52].

In this study, the infection rate of *E. bieneusi* was higher in the diarrheal foxes than it was in the non-diarrheal samples ( $p < 0.05$ ); a similar observation has been made in humans and cattle [36, 61]. Among patients exhibiting various degrees of diarrhea, the prevalence of *E. bieneusi* is notably higher compared to that among individuals without gastrointestinal (GI) disorders [3, 10, 60]. This may be because *E. bieneusi* can indeed cause diarrhea in the host, particularly under conditions of self-limiting infection or impaired host immunity [32]. Although the infection rate of *Blastocystis* sp. was higher in the diarrheal samples than it was in the non-diarrheal samples, the difference was not statistically significant ( $p > 0.05$ ).

Both *E. bieneusi* and *Blastocystis* sp. exhibited higher infection rates in juvenile foxes compared to those of the adult foxes. The findings of this study are consistent with prior research, indicating that juvenile foxes are more vulnerable to infection [8, 44]. Further epidemiological data and laboratory studies on their pathogenic mechanisms may be necessary to provide a scientific explanation for the differing infection rates of these two protozoa species in juvenile and adult foxes.

Eleven genotypes of *E. bieneusi* belonging to Group 1 were identified in the foxes in this study, including five known (D, Peru8, WildBoar 3, CHN-F1, and CHN-DC1) and six newly discovered genotypes (LNF-1, LNF-2, HBF-1, SDF-1, FJL-1, and FJL-2). Genotype D exhibits a strong capacity for cross-species transmission [14], and has been reported in humans in China, highlighting the need for vigilance regarding the potential transmission risk from foxes to humans [27, 35, 49, 50]. Additionally, the genotype Peru 8 has been reported to infect humans in other studies [2, 7]. Although only one Peru8-positive sample was found in this study, this genotype has been consistently detected in foxes, suggesting that they are susceptible hosts for genotype Peru 8 [62]. Notably, genotype CHN-F1 was predominant, found in all the subgroups, with a prevalence of 67.7% (115/170). First identified in dairy cattle in 2015, CHN-F1 has also been detected in raccoon dogs and has exhibited high prevalence rates in foxes in Henan Province (52.1%, 25/48) [44, 55, 66]. Aside from China, only two reports from Europe indicate that CHN-F1 can infect pigeons and chinchillas, suggesting its potential for global spread [22, 68]. The prevalence rates of WildBoar 3 and CHN-DC1 among the positive samples were 24.7% (42/170) and 0.6% (1/170), respectively. WildBoar 3 exhibits a broad spectrum of host adaptability and has been documented in wild boars from Poland, Czechia [30], and Spain [9], as well as in red foxes and stone martens from Portugal [13], and in goats from China [11]. Other reports confirm their presence in foxes, indicating their common occurrence [8, 62]. Phylogenetic

analysis identified six new genotypes of *E. bieneusi*. Group 1 is recognized for its zoonotic potential, indicating that these six novel genotypes may have the capacity for cross-host transmission [20]. Therefore, it is crucial to maintain ongoing attention to these genotypes in future investigations. Consequently, individuals such as fox breeders and those with weakened immunity – such as patients with HIV, children, and the elderly – should be especially vigilant regarding the risk of *E. bieneusi* transmission from foxes.

Only the ST3 subtype was detected in *Blastocystis* sp. in this study. This study was the first to identify the ST3 subtype in foxes. Previous studies have reported the presence of the ST1, ST4, and ST7 subtypes in foxes from northeast China [46], while foxes in Spain were found to harbor the ST7 and ST14 subtypes [6]. ST3 is the most common genotype in humans and the predominant subtype of *Blastocystis* sp. infections in China, with prevalence rates ranging from 43.8% to 62.0% [10, 31, 34, 37]. Another study has shown that the sequences of ST3 subtypes in dairy cows are identical to those found in exposed humans [17], suggesting a potential risk of ST3 cross-species transmission between humans and animals. It should be noted that there is 99.7% homology between the ST3 subtype (PQ223430) studied here and that in humans (MK782518). This indicates potential for the transmission of *Blastocystis* sp. between foxes and humans. However, additional follow-up studies are required to confirm this hypothesis. Other researchers should concurrently collect fecal samples from both feeders and foxes during the sampling process to determine whether such transmission is possible. ST3 is also widely prevalent among various animal species in China, such as rodents [48], wild rhesus macaques [56], Pallas's squirrels (*Callosciurus erythraeus*) [28], pet dogs [26], rabbits [58], pigs [19, 51], pigeons [43], raccoon dogs [46], and beef [47] and dairy cows [12, 41]. Given that ST3 is widely distributed in both humans and a variety of animals in China, it is important to continue investigating the ecological dynamics and transmission routes of *Blastocystis* sp.

This study investigated the prevalence of *E. bieneusi* genotypes and *Blastocystis* sp. subtypes in Chinese foxes, providing compelling evidence for the occurrence of infection in this particular host species. However, it is important to acknowledge certain limitations associated with this research. The sampling scope was confined to northeast China, potentially limiting the generalizability of the prevalence rates of *E. bieneusi* and *Blastocystis* sp. in foxes across China. Additionally, this study did not account for variations among fox breeds, which could influence the infection status and genetic distribution of these protists. Therefore, future research should broaden the sampling area and increase the sample size, while also meticulously documenting relevant sample information.

## Conclusion

There was a significant prevalence of *E. bieneusi* among foxes, with genotype CHN-F1 being widely distributed. Additionally, foxes carried genotypes D and Peru 8, which infect humans. Furthermore, new genotypes of *E. bieneusi* (LNF-1, LNF-2, SDF-1, HBF-1, FJL-1, and FJL-2) were identified for

the first time, all belonging to Group 1 with a potential zoonotic risk. ST3 was found to be the predominant subtype responsible for human *Blastocystis* sp. infection in China. Moreover, this study provides the first evidence that foxes serve as carriers of ST3. Consequently, foxes represent a plausible source of transmission for both *E. bienersi* and *Blastocystis* sp. infection. This investigation contributes to our understanding of the prevalence and genetic diversity of *E. bienersi* and *Blastocystis* sp. among the fox populations of China, while highlighting its public health significance in preventing potential zoonotic transmission mediated by foxes.

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### Conflicts of interest

The authors declare that they have no conflicts of interest.

### Data availability statement

The gene sequences obtained in this study have been submitted to GenBank (accession Nos. [PQ249072–PQ249088](#), and [PQ223428–PQ223432](#)).

### Author contribution statement

Yan Tang: Methodology, acquiring funding, software, investigation, visualization, and writing – original draft. Hai-Tao Wang: Methodology, software, investigation, visualization, and writing – original draft. Zhong-Yuan Li: Formal analysis and writing – review and editing. Jing Jiang: Formal analysis, resources, and writing – review and editing. Qing-Yu Hou and Xue-Min Li: Methodology and writing – review and editing. Li-Hua Yang: Conceptualization, data curation, methodology, supervision, and writing – review and editing. Ya Qin: Conceptualization, data curation, resources, and writing – original draft. All authors read and approved the final manuscript.

### References

- Abarca N, Santin M, Ortega S, Maloney JG, George NS, Molokin A, Cardona GA, Dashti A, Koster PC, Bailo B, Hernandez-de-Mingo M, Muadica AS, Calero-Bernal R, Carmena D, Gonzalez-Barrio D. 2021. Molecular detection and characterization of *Blastocystis* sp. and *Enterocytozoon bienersi* in cattle in northern Spain. *Veterinary Sciences*, 8, 191.
- Akinbo FO, Okaka CE, Omoregie R, Dearen T, Leon ET, Xiao L. 2012. Molecular epidemiologic characterization of *Enterocytozoon bienersi* in HIV-infected persons in Benin city, Nigeria. *American Journal of Tropical Medicine and Hygiene* 86, 441–445.
- Aydemir S, Halidi AG, Ekici A, Cengiz ZT. 2023. Investigation of the presence of *Enterocytozoon bienersi* and *Encephalitozoon intestinalis* in immunosuppressed patients with diarrhea by IFA and real time PCR methods. *Indian Journal of Medical Microbiology* 44, 100362.
- Aykur M, Malatyali E, Demirel F, Comert-Kocak B, Gentekaki E, Tsaousis AD, Dogruman-Al F. (2024). *Blastocystis*: a mysterious member of the gut microbiome. *Microorganisms*, 12, 461.
- Caballero-Gomez J, Avalos G, Matas-Mendez P, Figueiredo AM, Castro-Scholten S, Jimenez-Martin D, Koster PC, Santin M, Bailo B, Cano-Terriza D, Sarmiento P, Neves N, Carrapato C, Gonzalez-Barrio D, Mateo M, Garcia-Bocanegra I, Dashti A, Sanchez S, Carmena D. 2025. Dietary profiles of wild carnivores and *Blastocystis* occurrence: the case of the endangered Iberian lynx (*Lynx pardinus*) and systematic review. *Research in Veterinary Science* 184, 105518.
- Calero-Bernal R, Santin M, Maloney JG, Martin-Perez M, Habela MA, Fernandez-Garcia JL, Figueiredo A, Najera F, Palacios MJ, Mateo M, Balseiro A, Barral M, Lima-Barberoi JF, Koster PC, Carmena D. 2020. *Blastocystis* sp. subtype diversity in wild carnivore species from Spain. *Journal of Eukaryotic Microbiology* 67, 273–278.
- Chabchoub N, Abdelmalek R, Breton J, Kanoun F, Thellier M, Bouratbine A, Aoun K. 2012. Genotype identification of *Enterocytozoon bienersi* isolates from stool samples of HIV-infected tunisian patients. *Parasite*, 19, 147–151.
- Chen M, Wang H, Li X, Guo Y, Lu Y, Zheng L, Liang G, Sui Y, Wang B, Dai H, Dong H, Zhang L. 2024. Molecular epidemiology of *Enterocytozoon bienersi* from foxes and raccoon dogs in the Henan and Hebei Provinces in China. *BMC Veterinary Research*, 20, 53.
- Dashti A, Santin M, Koster PC, Bailo B, Ortega S, Imana E, Habela MA, Rivero-Juarez A, Vicente J, Arnal MC, de Luco DF, Morrondo P, Armenteros JA, Balseiro A, Cardona GA, Martinez-Carrasco C, Ortiz JA, Calero-Bernal R, Carmena D, Gonzalez-Barrio D. 2022. Zoonotic *Enterocytozoon bienersi* genotypes in free-ranging and farmed wild ungulates in Spain. *Medical Mycology*, 60, 9.
- Deng L, Chai Y, Zhou Z, Liu H, Zhong Z, Hu Y, Fu H, Yue C, Peng G. 2019. Epidemiology of *Blastocystis* sp. infection in China: a systematic review. *Parasite*, 26, 41.
- Dong H, Zhao Z, Zhao J, Fu Y, Lang J, Zhang J, Liang G, Zhang L, Li J, Zhao G. 2022. Molecular characterization and zoonotic potential of *Enterocytozoon bienersi* in ruminants in Northwest China. *Acta Tropica*, 234, 106622.
- Duan J, Qin H, Sun M, Fu Y, Lang J, Zhang A, Qin Z, Guo Z, Xu H, Li X, Wang C, Zhang L. 2024. Occurrence and genotypic identification of *Blastocystis* sp., *Enterocytozoon bienersi*, and *Giardia duodenalis* in dairy cattle in Heilongjiang Province, China. *Parasitology International*, 100, 102871.
- Figueiredo AM, Dashti A, Santin M, Koster PC, Torres RT, Fonseca C, Myserud A, Carvalho J, Sarmiento P, Neves N, Hipolito D, Palmeira JD, Teixeira D, Lima C, Calero-Bernal R, Carmena D. 2023. Occurrence and molecular characterization of *Enterocytozoon bienersi* in wild and domestic animal species in Portugal. *Medical Mycology*, 61, 2.
- Galvan-Diaz AL, Magnet A, Fenoy S, Henriques-Gil N, Haro M, Gordo FP, Millan J, Miro G, Del AC, Izquierdo F. 2014. Microsporidia detection and genotyping study of human pathogenic *E. bienersi* in animals from Spain. *Plos One*, 9, e92289.
- Gao ZQ, Wang HT, Hou QY, Qin Y, Qin SY, Zhao Q, Ma H. 2024. Wild rodents in three provinces of China exhibit a wide range of *Enterocytozoon bienersi* diversity. *Frontiers in Veterinary Science*, 11, 1427690.
- Gao ZQ, Wang HT, Hou QY, Qin Y, Yang X, Zhao Q, Ma H. 2024. Prevalence and subtypes of *Blastocystis* in wild rodents from three provinces in China. *Frontiers in Veterinary Science*, 11, 1432741.
- Greige S, El SD, Khaled S, Gantois N, Baydoun M, Chemaly M, Benamrouz-Vanneste S, Chabe M, Osman M, Certad G, Hamze M, Viscogliosi E. 2019. First report on the prevalence and subtype distribution of *Blastocystis* sp. in dairy cattle in Lebanon and assessment of zoonotic transmission. *Acta Tropica*, 194, 23–29.



18. Han B, Pan G, Weiss LM. 2021. Microsporidiosis in humans. *Clinical Microbiology Reviews* 34, e1020.
19. Han JQ, Li Z, Zou Y, Pu LH, Zhu XQ, Zou FC, Huang CQ. 2020. Prevalence, molecular characterization and risk factors of *Blastocystis* sp. from farmed pigs in Yunnan Province, south-western China. *Acta Parasitologica*, 65, 1005–1010.
20. Henriques-Gil N, Haro M, Izquierdo F, Fenoy S, Del AC. 2010. Phylogenetic approach to the variability of the microsporidian *Enterocytozoon bieneusi* and its implications for inter- and intrahost transmission. *Applied and Environmental Microbiology*, 76, 3333–3342.
21. Hernandez-Castro C, Maloney JG, Agudelo-Lopez SP, Toro-Londono MA, Botero-Garces JH, Orozco MC, Quintero-Quinchia YC, Correa-Cote JC, Munera-Duque A, Ricaurte-Ciro JC, Londono-Alvarez LI, Escobar RM, Koster PC, Sanchez S, Carmena D, Santin M. 2023. Identification and validation of novel *Blastocystis* subtype ST41 in a Colombian patient undergoing colorectal cancer screening. *Journal of Eukaryotic Microbiology*, 70, e12978.
22. Holubova N, Zikmundova V, Kicia M, Zajackowska Z, Rajsky M, Konecny R, Rost M, Mravcova K, Sak B, Kvac M. 2024. Genetic diversity of *Cryptosporidium* spp., *Encephalitozoon* spp. and *Enterocytozoon bieneusi* in feral and captive pigeons in central Europe. *Parasitology Research*, 123, 158.
23. Jiang S, Yu S, Feng Y, Zhang L, Santin M, Xiao L, Li W. 2024. Widespread distribution of human-infective *Enterocytozoon bieneusi* genotypes in small rodents in northeast China and phylogeny and zoonotic implications revisited. *Acta Tropica*, 253, 107160.
24. Jiang Y, Liu L, Yuan Z, Liu A, Cao J, Shen Y. 2023. Molecular identification and genetic characteristics of *Cryptosporidium* spp., *Giardia duodenalis*, and *Enterocytozoon bieneusi* in human immunodeficiency virus/acquired immunodeficiency syndrome patients in Shanghai, China. *Parasites & Vectors*, 16, 53.
25. Koehler AV, Herath H, Hall RS, Wilcox S, Gasser RB. 2024. Marked genetic diversity within *Blastocystis* in Australian wildlife revealed using a next generation sequencing-phylogenetic approach. *International Journal for Parasitology-Parasites and Wildlife*, 23, 100902.
26. Liao S, Lin X, Sun Y, Qi N, Lv M, Wu C, Li J, Hu J, Yu L, Cai H, Xiao W, Sun M, Li G. 2020. Occurrence and genotypes of *Cryptosporidium* spp., *Giardia duodenalis*, and *Blastocystis* sp. in household, shelter, breeding, and pet market dogs in Guangzhou, southern China. *Scientific Reports*, 10, 17736.
27. Liu H, Jiang Z, Yuan Z, Yin J, Wang Z, Yu B, Zhou D, Shen Y, Cao J. 2017. Infection by and genotype characteristics of *Enterocytozoon bieneusi* in HIV/AIDS patients from Guangxi Zhuang autonomous region, China. *BMC Infectious Diseases*, 17, 684.
28. Liu X, Ge Y, Wang R, Dong H, Yang X, Zhang L. 2021. First report of *Blastocystis* infection in Pallas's squirrels (*Callosciurus erythraeus*) in China. *Veterinary Research Communications*, 45, 441–445.
29. Maloney JG, Molokin A, Segui R, Maravilla P, Martinez-Hernandez F, Villalobos G, Tsaousis AD, Gentekaki E, Munoz-Antoli C, Klisiowicz DR, Oishi CY, Toledo R, Esteban JG, Koster PC, de Lucio A, Dashti A, Bailo B, Calero-Bernal R, Gonzalez-Barrio D, Carmena D, Santin M. 2022. Identification and molecular characterization of four new *Blastocystis* subtypes designated ST35-ST38. *Microorganisms*, 11, 46.
30. Nemejc K, Sak B, Kvetonova D, Hanzal V, Janiszewski P, Forejtek P, Rajsky D, Kotkova M, Ravaszova P, McEvoy J, Kvac M. 2014. Prevalence and diversity of *Encephalitozoon* spp. and *Enterocytozoon bieneusi* in wild boars (*Sus scrofa*) in central Europe. *Parasitology Research*, 113, 761–767.
31. Ning CQ, Ai L, Hu ZH, Chen JH, Tian LG. 2020. Progress of researches on *Blastocystis* infections in humans and animals in China. *Chinese Journal of Schistosomiasis Control*, 33, 95–101.
32. Nourrisson C, Lavergne RA, Moniot M, Morio F, Poirier P. 2024. *Enterocytozoon bieneusi*, a human pathogen. *Emerging Microbes & Infections*, 13, 2406276.
33. Piperni E, Nguyen LH, Manghi P, Kim H, Pasolli E, Andreu-Sanchez S, Arre A, Birmingham KM, Blanco-Miguez A, Manara S, Valles-Colomer M, Bakker E, Busonero F, Davies R, Fiorillo E, Giordano F, Hadjigeorgiou G, Leeming ER, Lobina M, Masala M, Maschio A, McIver LJ, Pala M, Pitzalis M, Wolf J, Fu J, Zhernakova A, Caccio SM, Cucca F, Berry SE, Ercolini D, Chan AT, Huttenhower C, Spector TD, Segata N, Asnicar F. 2024. Intestinal *Blastocystis* is linked to healthier diets and more favorable cardiometabolic outcomes in 56, 989 individuals from 32 countries. *Cell*, 187, 4554–4570.
34. Poulsen CS, Efunshile AM, Nelson JA, Stensvold CR. 2016. Epidemiological aspects of *Blastocystis* colonization in children in Ilero, Nigeria. *American Journal of Tropical Medicine and Hygiene*, 95, 175–179.
35. Qi M, Yu F, Zhao A, Zhang Y, Wei Z, Li D, Zhang L. 2020. Unusual dominant genotype NIA1 of *Enterocytozoon bieneusi* in children in southern Xinjiang, China. *PLoS Neglected Tropical Diseases*, 14, e8293.
36. Qin Y, Chen C, Qin Y, Yang X, Li M, Meng X, Zhao Z, Ma N, Cai Y, Zhang Y, Zhao Q. 2022. Prevalence and related factors of *Enterocytozoon bieneusi* in cattle: a global systematic review and meta-analysis. *Preventive Veterinary Medicine*, 208, 105775.
37. Rauff-Adedotun AA, Meor TF, Shaari N, Lee IL. 2021. The coexistence of *Blastocystis* spp. in humans, animals and environmental sources from 2010–2021 in Asia. *Biology*, 10, 990.
38. Santin M, Fayer R. 2009. *Enterocytozoon bieneusi* genotype nomenclature based on the internal transcribed spacer sequence: a consensus. *Journal of Eukaryotic Microbiology*, 56, 34, 38.
39. Sejnohova A, Koutenska M, Jirku M, Brozova K, Pavlickova Z, Kadlecova O, Cinek O, Maloney JG, Santin M, Petrzalkova KJ, Jirku K. 2024. A cross-sectional survey of *Blastocystis* sp. and *Dientamoeba fragilis* in non-human primates and their caregivers in Czech zoos. *One Health*, 19, 100862.
40. Silberman JD, Sogin ML, Leipe DD, Clark CG. 1996. Human parasite finds taxonomic home. *Nature*, 380, 398.
41. Song JK, Yin YL, Yuan YJ, Tang H, Ren GJ, Zhang HJ, Li ZX, Zhang YM, Zhao GH. 2017. First genotyping of *Blastocystis* sp. in dairy, meat, and cashmere goats in northwestern China. *Acta Tropica*, 176, 277–282.
42. Stensvold CR, Tan K, Clark CG. 2020. *Blastocystis*. *Trends in Parasitology*, 36, 315–316.
43. Su C, Mei X, Wei L, Wang J, Feng X, Wang P, He B, Chang Y, Xu F, Wang M, Tian X, Zhang Z, Li X, Wang S. 2022. Prevalence and molecular subtyping of *Blastocystis* in domestic pigeons in Henan province, central China. *Journal of Eukaryotic Microbiology*, 69, e12888.
44. Sui Y, Tong C, Li X, Zheng L, Guo Y, Lu Y, Huang S, Wang H, Chen M, Xu C, Dai H, Liu F, Dong H, Zhang L. 2021. Molecular detection and genotyping of *Enterocytozoon bieneusi* in captive foxes in Xinxiang, central China and its impact on gut bacterial communities. *Research in Veterinary Science*, 141, 138–144.
45. Tomiak J, Stensvold CR. 2024. Accelerating the paradigm shift in *Blastocystis* research. *Trends in Parasitology*, 40, 775–776.

46. Wang J, Gong B, Liu X, Zhao W, Bu T, Zhang W, Liu A, Yang F. 2018. Distribution and genetic diversity of *Blastocystis* subtypes in various mammal and bird species in northeastern China. *Parasites & Vectors*, 11, 522.
47. Wang J, Gong B, Yang F, Zhang W, Zheng Y, Liu A. 2018. Subtype distribution and genetic characterizations of *Blastocystis* in pigs, cattle, sheep and goats in northeastern China's Heilongjiang Province. *Infection Genetics and Evolution*, 57, 171–176.
48. Wang J, Wang Y, Huang W, Zhang T, Yu K, Chen J, Zhou L, Cao W, Xu J, Ma J, Huang H, Zhao W. 2024. Molecular prevalence, subtype distribution, and zoonotic potential of *Blastocystis* sp. in wild rodents and shrews inhabiting Zhejiang Province of China. *Frontiers in Veterinary Science*, 11, 1427490.
49. Wang L, Xiao L, Duan L, Ye J, Guo Y, Guo M, Liu L, Feng Y. 2013. Concurrent infections of *Giardia duodenalis*, *Enterocytozoon bieneusi*, and *Clostridium difficile* in children during a *Cryptosporidiosis* outbreak in a pediatric hospital in China. *PLoS Neglected Tropical Diseases*, 7, e2437.
50. Wang L, Zhang H, Zhao X, Zhang L, Zhang G, Guo M, Liu L, Feng Y, Xiao L. 2013. Zoonotic cryptosporidium species and *Enterocytozoon bieneusi* genotypes in HIV-positive patients on antiretroviral therapy. *Journal of Clinical Microbiology*, 51, 557–563.
51. Wang P, Li S, Zou Y, Hong ZW, Wang P, Zhu XQ, Song DP, Chen XQ. 2021. Prevalence and subtype distribution of *Blastocystis* sp. in diarrheic pigs in southern China. *Pathogens*, 10, 1189.
52. Wang S, Bai X, Zhang X, Reis S, Chen D, Xu J, Gu B. 2021. Urbanization can benefit agricultural production with large-scale farming in China. *Nature Food*, 2, 183–191.
53. Wang SS, Wang RJ, Fan XC, Liu TL, Zhang LX, Zhao GH. 2018. Prevalence and genotypes of *Enterocytozoon bieneusi* in China. *Acta Tropica*, 183, 142–152.
54. Xue NY, Li ZY, Wang HT, Qin Y, Li XM, Hou QY, Jiang J, Yang X, Ni HB. 2024. High genetic diversity of *Enterocytozoon bieneusi* in minks and raccoon dogs in northern China. *Parasite*, 31, 71.
55. Yang Y, Lin Y, Li Q, Zhang S, Tao W, Wan Q, Jiang Y, Li W. 2015. Widespread presence of human-pathogenic *Enterocytozoon bieneusi* genotype D in farmed foxes (*Vulpes vulpes*) and raccoon dogs (*Nyctereutes procyonoides*) in China: first identification and zoonotic concern. *Parasitology Research*, 114, 4341–4348.
56. Yu M, Yao Y, Xiao H, Xie M, Xiong Y, Yang S, Ni Q, Zhang M, Xu H. 2023. Extensive prevalence and significant genetic differentiation of *Blastocystis* in high- and low-altitude populations of wild rhesus macaques in China. *Parasites & Vectors*, 16, 107.
57. Yu X, Wang H, Li Y, Mu X, Yuan K, Wu A, Guo J, Hong Y, Zhang H. 2023. Occurrence and genotypic identification of *Blastocystis* spp., *Enterocytozoon bieneusi*, and *Giardia duodenalis* in Leizhou black goats in Zhanjiang city, Guangdong Province, China. *Animals*, 13, 2777.
58. Zhang M, Zhang J, Sun J, Zhu S, Xu H, Li X, Zhang L, Li J. 2023. First identification of *Blastocystis* sp. subtypes in Rex rabbits in China. *Parasitology Research*, 122, 1079–1086.
59. Zhang P, Zhang Q, Han S, Yuan G, Bai J, He H. 2022. Occurrence and genetic diversity of the zoonotic enteric protozoans and *Enterocytozoon bieneusi* in Pere David's deer (*Elaphurus davidianus*) from Beijing, China. *Pathogens*, 11, 1223.
60. Zhang SX, Zhou YM, Xu W, Tian LG, Chen JX, Chen SH, Dang ZS, Gu WP, Yin JW, Serrano E, Zhou XN. 2016. Impact of co-infections with enteric pathogens on children suffering from acute diarrhea in southwest China. *Infectious Diseases of Poverty*, 5, 64.
61. Zhang W, Ren G, Zhao W, Yang Z, Shen Y, Sun Y, Liu A, Cao J. 2017. Genotyping of *Enterocytozoon bieneusi* and subtyping of *Blastocystis* in cancer patients: relationship to diarrhea and assessment of zoonotic transmission. *Frontiers in Microbiology*, 8, 1835.
62. Zhang XX, Cong W, Lou ZL, Ma JG, Zheng WB, Yao QX, Zhao Q, Zhu XQ. 2016. Prevalence, risk factors and multilocus genotyping of *Enterocytozoon bieneusi* in farmed foxes (*Vulpes lagopus*), northern China. *Parasites & Vectors*, 9, 72.
63. Zhang XX, Tan QD, Zhou DH, Ni XT, Liu GX, Yang YC, Zhu XQ. 2015. Prevalence and molecular characterization of *Cryptosporidium* spp. in dairy cattle, northwest China. *Parasitology Research*, 114, 2781–2787.
64. Zhang Y, Xin L, Zhao A, Xu C, Wang T, Jing B, Qi M. 2021. Molecular detection and genotypes of *Enterocytozoon bieneusi* in farmed mink (*Neovison vison*), blue foxes (*Alopex lagopus*), and raccoon dogs (*Nyctereutes procyonoides*) in Xinjiang, China. *International Journal for Parasitology-Parasites and Wildlife*, 14, 211–215.
65. Zhao W, Ren G, Wang L, Xie L, Wang J, Mao J, Sun Y, Lu G, Huang H. 2024. Molecular prevalence and subtype distribution of *Blastocystis* spp. among children who have diarrhea or are asymptomatic in Wenzhou, Zhejiang Province, China. *Parasite*, 31, 12.
66. Zhao W, Zhang W, Yang Z, Liu A, Zhang L, Yang F, Wang R, Ling H. 2015. Genotyping of *Enterocytozoon bieneusi* in farmed blue foxes (*Alopex lagopus*) and raccoon dogs (*Nyctereutes procyonoides*) in China. *PLoS One*, 10, e142611.
67. Zhao W, Zhang Y, Li J, Ren G, Qiang Y, Wang Y, Lai X, Lei S, Liu R, Chen Y, Huang H, Li W, Lu G, Tan F. 2023. Prevalence and distribution of subtypes of *Blastocystis* in Asiatic brush-tailed porcupines (*Atherurus macrourus*), bamboo rats (*Rhizomys pruinosus*), and masked palm civets (*Paguma larvata*) farmed in Hainan, China. *Parasite*, 30, 45.
68. Zikmundova V, Horakova V, Tumova L, Koudela B, Holubova N, Sak B, Rost M, Beranova K, Kvac M. 2024. Pet chinchillas (*Chinchilla lanigera*): source of zoonotic *Giardia intestinalis*, *Cryptosporidium ubiquitum* and microsporidia of the genera *Encephalitozoon* and *Enterocytozoon*. *Veterinary Parasitology*, 331, 110275.



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