

GROWTH-PROMOTING EFFECT ON IRON-SULFUR PROTEINS ON AXENIC CULTURES OF *ENTAMOEBA DISPAR*

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Summary:

A growth-promoting factor (GPF) that promotes the growth of *Entamoeba dispar* under axenic culture conditions was found in fractions of mitochondria (Mt), hydrogenosomes (Hg) and chloroplasts (Cp) obtained from cells of six different protozoan, mammalian and plant species. We were able to extract the GPF from the Cp-rich leaf cells of a plant (spiderwort: *Commelinia communis* L.) in an acetone-soluble fraction as a complex of chlorophyll with low molecular weight proteins (molecular weight [MW] approximately 4,600). We also found that on treatment with 0.6 % complexes of 2-mercaptoproethanol (2ME), complexes of chlorophyll-a with iron-sulphur (Fe-S) proteins (e.g., ferredoxins [Fd] from spinach and *Clostridium pasteurianum*) and noncomplex rubredoxin (Rd) from *C. pasteurianum* have a growth-promoting effect on *E. dispar*. These findings suggest that *E. dispar* may lack a sufficient quantity of some essential components of Fe-S proteins, such as Fe-S center.

KEY WORDS : growth, mitochondria, hydrogenosomes, chloroplasts, iron-sulphur protein, *Entamoeba dispar*.

Résumé : LES EFFETS D'ACCÉLÉRATION DE CROISSANCE DES PROTÉINES FER-SOUFRE DANS LA CULTURE AXÉNIQUE D'*ENTAMOEBA DISPAR*

Des facteurs d'accélération de croissance (*Growth promoting factor: GPF*) favorisant le développement d'*Entamoeba dispar* ont été détectés dans la composition de la mitochondrie, de l'hydrogénosome et du chloroplaste isolés à partir de six sortes de cellules issues de protozoaires, mammifères et plantes. De plus, les GPF de cellules de mésophile contenant une grande quantité de chloroplaste végétal (*Commelinia communis* L.) ont pu être extraits comme une substance composée de protéines de faible masse moléculaire ($\approx 4\,600$) et de chloroplastes dans la composition acétone-soluble. À partir de ces résultats, nous avons préalablement pu découvrir des effets d'accélération de croissance dans le corps composé de protéines fer-soufre (épinard et ferredoxine de *Clostridium pasteurianum*) et de chlorophylle-a traité au 2-mercaptoproéthanol 0,6 % et du corps simple rubrédoxine de *C. pasteurianum*. Ces observations ont suggéré qu'une composante essentielle formant la protéine fer-soufre d'*E. dispar* (comme noyau fer-soufre) semblait être insuffisante.

MOTS CLÉS : croissance, mitochondrie, hydrogénosome, chloroplaste, protéine fer-soufre, *Entamoeba dispar*.

INTRODUCTION

Entamoeba dispar grows well under xenic and monoxenic culture conditions along with enteric bacteria such as *Escherichia coli* or anaerobic bacteria such as *Fusobacterium symbiosum* (*Clostridium symbiosum* ATCC 14940) (Robinson, 1968; Diamond, 1982; Vargas *et al.*, 1990). However, even after the introduction of the axenic yeast extract-iron serum (YI-S) medium for *E. dispar* (Diamond *et al.*, 1995; Clark, 1995) the axenic cultivation of *E. dispar* remains difficult. Compared with *E. dispar*, pathogenic *E. histolytica*, which is closely related to *E. dispar*, easily adapts to the axenic culture medium (TYI-S-33) (Diamond *et*

al., 1978). Additionally, *Entamoeba histolytica* is capable of invading the mucosa of the large intestine. We have also developed a new yeast extract-iron-gluconic acid-dihydroxyacetone-serum medium (YIGADHA-S) (Kobayashi *et al.*, 2005) based on the YI-S medium and on the results of an investigation on bacterial metabolic products, and have succeeded in culturing five strains under axenic conditions. However, despite using the YIGADHA-S culture system, the axenic growth of four of five strains of *E. dispar* was found to be very poor, with the exception of one primate-derived strain (CYNO 09: TPC) isolated from a cynomolgus monkey. A further search for useful growth promoting factors (GPFs) revealed that autoclaved (121° C, 15 minutes) bacteria and more than 20 types of protozoan, mammalian and plant cells containing mitochondria (Mt), hydrogenosomes (Hg) and chloroplasts (Cp) have a significant growth-promoting effect on *E. dispar*. However, the degree of these growth-promoting effects differs among the GPFs. In the present study, we were able to extract GPF from the Cp-rich leaf cells of a plant (spiderwort: *Commelinia communis* L.) in an acetone-soluble fraction as a chlorophyll complex with low mole-

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cular weight proteins. Significant amounts of iron (Fe), sulphur (S) and molybdenum (Mo) atoms were found in the protein-chlorophyll complex fraction along with the magnesium (Mg) atom of chlorophyll.

Based on these findings, we hypothesized that some common components of Fe-S proteins present in Mt, Hg, Cp and bacteria support the growth of *E. dispar*. This is because in *E. dispar*, an essential redox Fe-S protein [ferredoxin (Fd)] is expected to be involved in energy metabolism, such as oxidative decarboxylation of pyruvate to acetyl-coenzyme A (acetyl-CoA), which is similar to that found in *E. histolytica* (McLaughlin & Aley, 1985).

MATERIALS AND METHODS

E. DISPAR ISOLATE

In this study, one human-derived strain of axenically grown *E. dispar* (AS 16 IR) (Kobayashi *et al.*, 2005) was used to determine the growth promoting effect of Fe-S proteins.

AXENIC CULTIVATION OF *E. DISPAR*

The YIGADHA-S medium (Kobayashi *et al.*, 2005) containing 15 % heat-inactivated bovine serum was used as the axenic cultivation medium for *E. dispar*.

DONOR CELLS OF MT, HG, CP AND MITOSOMES

The Mt donors are as follows: protozoan parasites: i) *Cryptosporidium fasciculata* (ReF-1, PRR strain, ATCC 50083); ii) *Trypanosoma cruzi* (Tulahuen strain); iii) *Leishmania major* (MHOM/SU/73/5-ASKH strain, ATCC 50155); and iv) *Acanthamoeba castellanii* (ATCC 30011). v) The vertebrate cells used as Mt donors were from the mouse lymphoblast cell line (P388D1, ATCC CCL-46). The Hg donors are as follows: protozoan parasites: vi) *Trichomonas vaginalis* (KO-11 strain) (Qi *et al.*, 1995) and vii) *Trichomonas foetus* (Okamoto *et al.*, 1998). viii) Leaves of the spiderwort (*Commelinaceae communis* L.) were used as the Cp donors. Consisting amitochondrial protozoan parasite mitosomes that are mitochondrial remnant organelles (Tovar *et al.*, 2003) was used as the mitosome donor, ix) *Giardia intestinalis* (syn. *lamblia*), (Portland-I strain, ATCC 30888). The bacterium used was *Pseudomonas aeruginosa* (PA:KEIO strain) (Kobayashi *et al.*, 1998).

PREPARATION OF CELLS

C. fasciculata and *T. cruzi* were axenically cultured in liver infusion tryptose (LIT) medium (Gutteridge *et al.*, 1969) supplemented with 10 % foetal bovine serum (FBS). After cultivation for three and seven days res-

pectively, 2×10^7 cells of each parasite were harvested by centrifugation (650 g \times 8 minutes). The cells of each parasite were washed three times by centrifugation with 10 mM phosphate-buffered saline (PBS) (pH 7.4), and each pellet was suspended in approximately eight times its volume of 50 mM Tris-HCl (pH 7.4). *L. major* was axenically cultured in Schneider's medium (GibcoTM) supplemented with 15 % FBS at 26°C for four days and harvested by centrifugation (650 g \times 8 minutes). *A. castellanii* was axenically cultured in peptone-yeast glucose (PYG) medium (Rowbotham, 1983; ATCC media formulations No. 712) for four days and harvested by centrifugation (650 g \times 4 minutes). The P388D1 cell line was cultured in RPMI 1640 medium (Nissui Pharmaceutical Co., Taito-ku, Tokyo, Japan) supplemented with 10 % FBS for four days and harvested by centrifugation (125 g \times 4 minutes). *T. vaginalis* and *T. foetus* were axenically cultured for three days in BI-S-33 medium in which the peptone components of TYI-S-33 medium (Diamond *et al.*, 1978) are substituted by Biosate (BBL, Becton Dickinson Co., Cockeysville, Maryland, USA), and harvested by centrifugation (275 g \times 4 minutes). The Cp-rich fraction from the wild spiderwort (*C. communis* L.) was isolated. The leaves of wild spiderwort were picked from the private field of Keio University (Tokyo) during the flower season from June to August (2000-2003). Fresh green spiderwort leaves were stripped from the stems, washed with tap water and then with distilled water. The leaves were ground with serum-free RPMI 1640 medium (RPMI) in an earthenware mortar by using a wooden pestle. The resulting leaf cell suspension was filtered through a double thickness gauze to remove the residue. The cell suspension was then washed three times with RPMI by centrifugation (440 g \times 10 minutes). *P. aeruginosa* was cultured in Biosate-Iron broth (Diamond *et al.*, 1978) for three days and harvested by centrifugation (650 g \times 4 minutes). *G. intestinalis* was axenically cultured in modified BI-S-33 medium (Keister, 1983) for three days and harvested by centrifugation (275 g \times 4 minutes).

PREPARATION OF MT, HG AND CP FRACTIONS

Mt, Hg and Cp rich fractions were prepared as described by Hogeboom (1955), Opperdoes *et al.* (1984) and Gorham (1955), respectively. Since the methodology for mitosome isolation has not been established, intact cells of *G. intestinalis* were tested for their growth-promoting effect without preparing the mitosome rich fraction. After washing the fractions with RPMI once by centrifugation (440 g \times 10 minutes), each fraction was suspended in 2 ml of RPMI. In order to test for their growth-promoting effect under axenic culture conditions, the fractions were sterilized by autoclaving (121°C, 15 minutes), and 0.2 ml of each suspension was added into the YIGADHA-S medium (5 ml). The

medium was then inoculated with a 0.8 ml suspension of *E. dispar* (AS16IR strain = AS 16 IR) (final density: 200-2,700 amoebae/ml).

NuPAGE® NOVEX 4-12 % BIS-TRIS GRADIENT PEPTIDE GEL ELECTROPHORESIS AND PREPARATIVE SODIUM DODECYL SULFATE-POLYACRYLAMIDE GEL ELECTROPHORESIS (SDS-PAGE)

The lyophilized cell fraction of spiderwort was extracted with 20 %, 40 %, 80 % and 100 % acetone solutions (20 mg/ml) for 10 minutes at 4°C. After centrifugation (17,800 g × 10 minutes, 4°C) of each acetone extract, the supernatant was collected. The supernatants that were extracted with 80 % and 100 % acetone solutions were diluted with distilled water to obtain an acetone concentration of 50 %. Each supernatant was then concentrated by evaporation by using a SpeedVac system (SVC 100; Savant Instruments, Inc., Farmingdale, New York, USA) until evaporated to dryness. The molecular weight of the proteins in each extract was determined by NuPAGE® Novex 4-12 % Bis-Tris gradient peptide (Invitrogen Corp., Carlsbad, California, USA) gel electrophoresis.

The bands of complexes of low molecular weight protein with chlorophyll that were detected in the supernatants of 80 % and 100 % acetone extracts were isolated by preparative SDS-PAGE on a 12 % gel in a Mini Prep Cell (Bio-Rad Laboratories, Inc., Hercules, California, USA) electrophoresis unit. SDS-PAGE was performed according to the procedure described by Laemmli (1970). For NuPAGE® (Novex 4-12 % gradient peptide gel = Novex 4-12 %) each dried and concentrated supernatant of the acetone extract was redissolved in the sample buffer with 50 mM dithiothreitol and heated at 70°C for 10 minutes to reduce the disulfide bonds. For SDS-PAGE (12 % gel) each supernatant was redissolved in the sample buffer with 1 % (v/v) 2-mercaptoethanol (2ME) and boiled for four minutes. Following NuPAGE®, the gel was stained with 2D-silverstain II (Daiichi Pure Chemicals Co. Ltd., Chuo-ku, Tokyo, Japan).

ASSAY OF THE ACETONE EXTRACTS OF SPIDERWORT LEAF CELLS FOR A GPF

One millilitre supernatants (obtained after centrifugation; 17,800 g × 10 minutes, 4°C) of the 20 %, 40 %, 80 % and 100 % acetone extracts of the lyophilized cell fraction of spiderwort (20 mg/ml) was dialyzed three times against 200-250 volumes of distilled water for 18 hours by using a Spectra/Por® 3 (3500 MWCO) dialysis membrane for 18 h; the volume of each extract was adjusted to 2.4 ml. Each extract was sterilized by filtration (Sartorius membrane filter; 0.2-μm pore size), and 0.2 ml of each extract was added to the YIGADHA-S medium to test the growth-promoting effect of the extract on *E. dispar* (AS 16 IR).

ANALYSIS USING A SCANNING ELECTRON MICROSCOPY (SEM)/ENERGY DISPERITIVE X-RAY ANALYSER (EDX)-INTEGRATED ANALYSIS SYSTEM

Following the preparative SDS-PAGE, the complexes of chlorophyll with low molecular weight proteins in the 80 % and 100 % acetone-soluble fractions were dialyzed in the same manner as described above. After lyophilization, the dialyzed complexes were examined using an SEM and EDX-integrated analysis system SEM-EDX III Type N/H (Hitachi Science Systems, Ltd., Hitachinaka, Ibaragi, Japan) in the Nihonbashi laboratory, Hitachi High-Technologies Co., Tokyo, Japan.

PREPARATION OF CHLOROPHYLL-A COMPLEXES WITH PURIFIED IRON-SULPHUR PROTEINS

Purified Fd from spinach (1 μg/μl; Sigma F-3013) and *C. pasteurianum* (1 μg/μl; Sigma F-7629) and purified rubredoxin (Rd) from *C. pasteurianum* (15.7 μg/μl; Sigma R-2512) were dissolved in 0.6 % 2ME/H₂O (20 μg of Fd or 31.4 μg of Rd/4 ml of 0.6 % 2ME), and the Fd and Rd solutions were dialyzed in the same manner as described above. Purified chlorophyll-a (Sigma C-5753) was dissolved in acetone (25 μg/25 μl acetone), and after adding 4 ml of distilled water, the solution was dialyzed in the same manner as described above. The Fd and Rd solutions were mixed with dialyzed chlorophyll-a and incubated for 10 minutes at 25°C. The solutions of chlorophyll-a complexes were then sterilized by filtration, and 0.2 ml of each solution was added to the YIGADHA-S medium to test their growth-promoting effect on *E. dispar* AS 16 IR.

STATISTICAL ANALYSIS OF GROWTH-PROMOTING EFFECT ON *E. DISPAR*

All experiments for testing growth-promoting effect were repeated at least twice. The data of each experiment were subjected to one-way analysis of variance (ANOVA) with Dunnett's multiple comparison post hoc test; the levels of statistical significance were taken as *p* < 0.05 and *p* < 0.01.

RESULTS

The Mt, Hg and Cp isolated from eight different types of cells were tested to determine whether they promoted the growth of axenically grown *E. dispar* AS 16 IR. The intact cells of *G. intestinalis* and *P. aeruginosa* were also tested for the same. A growth-promoting effect was apparently observed for every fraction except for the fraction of intact cells of *G. intestinalis*. Apparently, the intact cells of *E. histolytica* (HM-1: IMSS clone 6 strain) containing mitosomes (Leon-Avila & Tovar, 2004) also did not produce a

growth-promoting effect (data not shown). Although the growth-promoting effect of the two fractions of Mt from *T. cruzi* and the Hg fraction from *T. vaginalis* retained the statistically significant difference by Dunnett's test ($p < 0.05$), the growth-promoting effect of the other six fractions containing Mt, Hg and Cp was significantly different ($p < 0.01$ or $p < 0.05$). In particular, the effect of the Cp fraction from spiderwort exceeded that of the intact cells fraction of *P. aeruginosa* as shown in Figure 1. Thus, it was concluded that some of the Mt, Hg and Cp fractions, which were obtained under appropriate conditions as well as the intact bacterial cells contain a GPF.

Acetone extraction was used to obtain a water-soluble GPF from Cp-rich leaf cells of some plant species (e.g., spiderwort, cherry and morning glory), which have sufficiently strong leaf-cell membranes that can withstand the leaf crushing and cell isolation process. Figure 2 shows the results obtained on testing the 0 %, 20 %, 40 %, 80 % and 100 % acetone-soluble fractions from lyophilized Cp-rich leaf cells of spiderwort for a growth-promoting effect. The results showed that the 20 % and 80 % acetone-soluble fractions had a stronger growth-promoting effect than the other acetone-soluble fractions (0 %, 40 % and 100 %). The growth-promoting effects of the 20 % and 80 % acetone-soluble fractions were statistically significant at $p < 0.01$ (Dunnett's test);

however, the effects of the 0 %, 40 % and 100 % acetone-soluble fractions were retained at $p < 0.05$. The silver-stain NuPAGE® analysis of these acetone-soluble fractions yielded a distinct dense protein band (MW 4600) that was common to the 20 %, 40 %, 80 % and 100 % acetone-soluble fractions. The intensity of the bands of the 20 % and 80 % acetone-soluble fractions were stronger than the bands of the other acetone-soluble fractions (40 % and 100 %) (Fig. 3). These results indicated that the efficacy of the growth-promoting effect was correlated with the intensity of the low molecular weight protein band (Figs 2, 3). The band of the complex of the chlorophyll with low molecular weight proteins (green colour) present in both the 80 % and 100 % acetone-soluble fractions was obtained by SDS-preparative electrophoresis; both the fractions showed a growth-promoting effect on *E. dispar* AS 16 IR (data not shown). The result showed that the band of the complex of chlorophyll with low molecular weight proteins, which was present in the acetone-soluble fractions, contained a GPF. The reduction in the intensity of the low molecular weight protein bands obtained by using the 40 % acetone-soluble fraction was considered to be caused by the incomplete solubility of chlorophyll. Some protein parts are believed to be precipitated together with the insoluble chlorophyll when extraction is performed with an intermediate concen-

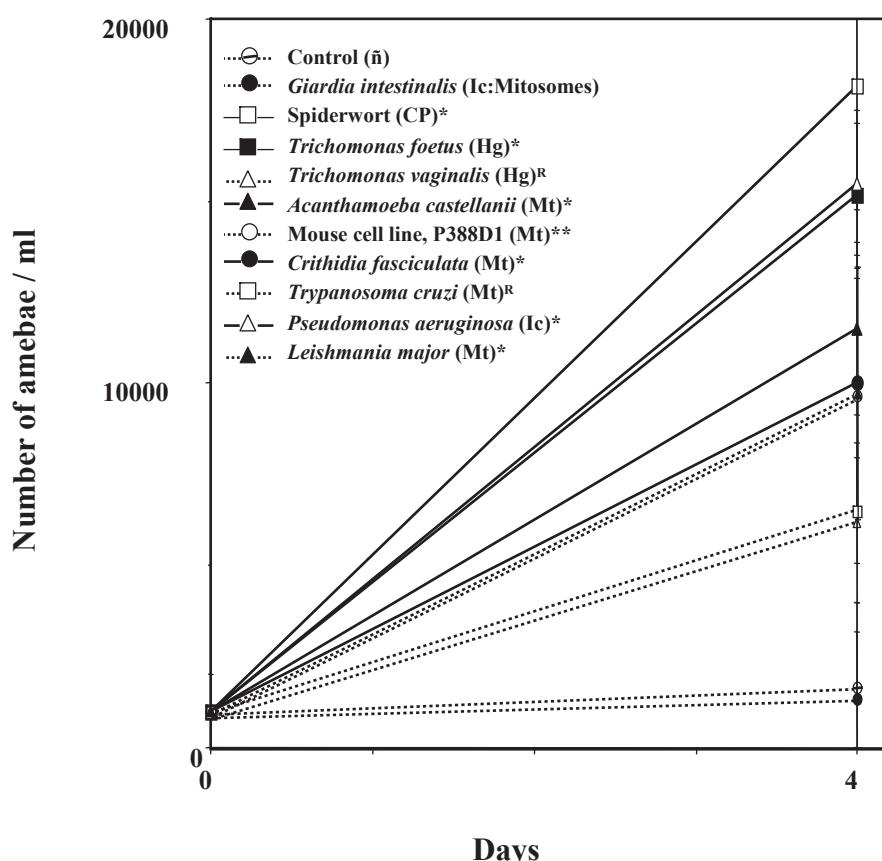


Fig. 1. – The effect of intact cells (Ic) and the fractions of mitochondria (Mt), chloroplasts (Cp) and hydrogenosomes (Hg) from ten types of cells, including bacterial, mammalian, plant and protozoan cells, on the growth of *Entamoeba dispar*.

The cellular components were sterilized by autoclaving at 121°C for 15 minutes. The growth kinetics of the *E. dispar* AS 16 IR strain in the YIGADHA-S medium are shown (mean numbers of amoebae in duplicate cultures are plotted). *, **: the mean of the growth-kinetic level was significantly higher than that of the control (* $p < 0.01$ and ** $p < 0.05$ by Dunnett's test). R: the significant difference was retained by Dunnett's test ($p < 0.05$).

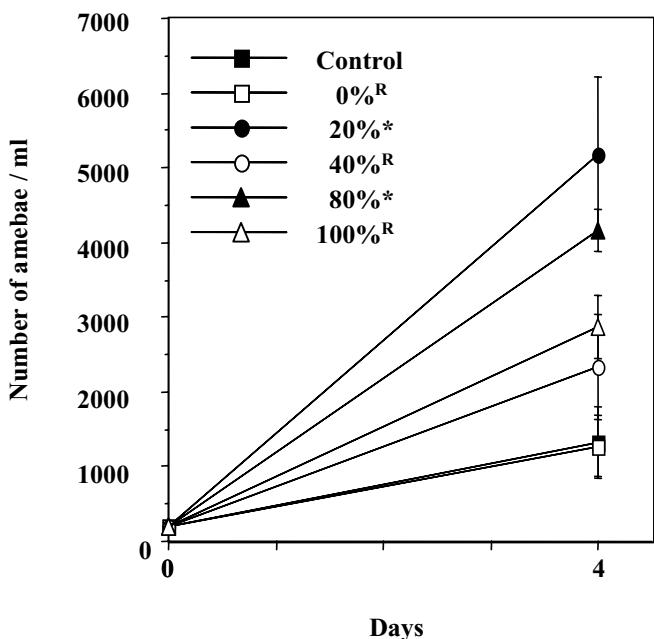


Fig. 2. – Effect of soluble fractions of spiderwort Cp-rich leaf cells extracted with five different concentrations of acetone (0 %, 20 %, 40 %, 80 % and 100 %) on the growth of *E. dispar*.

The growth kinetics of the *E. dispar* AS 16 IR strain in the YIGADHA-S medium are shown (mean numbers of amoebae in duplicate cultures are plotted). *: the mean of the growth-kinetic level was significantly higher than that of the control (* $p < 0.01$ by Dunnett's test). R: the significant difference was retained by Dunnett's test ($p < 0.05$).

tration of acetone such as 40 %. As compared to an 80 % acetone concentration, the 100 % concentration of acetone was considered to be slightly severe for the extraction of the low molecular weight proteins without

decreasing the activity of a GPF. As a result, the intensity of the low molecular weight protein bands obtained by using the 40 % and 100 % acetone-soluble fraction was believed to be reduced.

The SEM/EDX-integrated analysis system revealed that the low molecular weight protein band contained Fe, S and Mo atoms along with the Mg atom of chlorophyll (Figs 4, 5). We believed that Fd present in Cp is a type of Fe-S protein containing Fe and S atoms. In order to confirm the growth-promoting effect of Fd, we examined the effects of purified spinach Fd (MW 12,000), *C. pasteurianum* Fd (MW 6,000) and purified *C. pasteurianum* Rd (MW 19,000) on the growth of *E. dispar* AS 16 IR. The analysis revealed that 0.6 % 2ME-treated *C. pasteurianum* Rd and each of the water-soluble chlorophyll-a complexes with 0.6 % 2ME-treated spinach Fd and *C. pasteurianum* Fd have a growth-promoting effect on *E. dispar* AS 16 IR with a statistically significant difference ($p < 0.01$) (Fig. 6). The non-complexed Fd from spinach and from *C. pasteurianum* treated with 0.6 % 2ME also had a slight growth-promoting effect when compared with the growth kinetics of the control with or without chlorophyll-a; however, these growth-promoting effects were retained ($p < 0.05$).

These results suggest that the GPF in the acetone-soluble fraction of spiderwort Cp-rich leaf cells is the complex of the chlorophyll-a with a component of an Fe-S protein, such as an inorganic Fe-S centre, released by the breakage of disulfide bonds due to treatment with 2ME. The Fe-S redox proteins are commonly found in enteric bacteria (e.g., Fd), mitochondria (e.g., Fe-S proteins in complex I and II), chloroplasts (e.g., Fd) and hydrogenosomes (e.g., Fd); however, there are diffe-

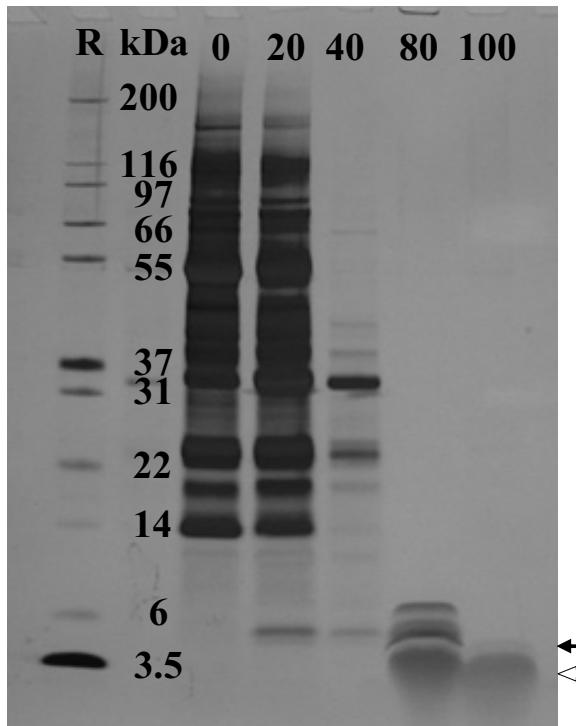


Fig. 3. – NuPAGE® (Novex 4-12 %) of five acetone-soluble fractions of spiderwort Cp-rich leaf cells extracted with different concentrations of acetone (0 %, 20 %, 40 %, 80 % and 100 %).

Molecular mass markers (M) are shown on the left. ← : acetone-soluble low molecular weight protein bands were isolated from the 20 %, 40 %, 80 % and 100 % acetone-soluble fractions. ⇝: broad green band of chlorophyll.

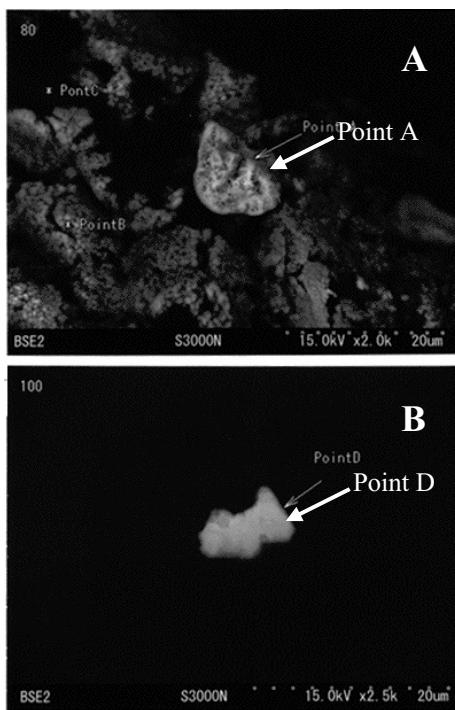


Fig. 4. – Scanning electron microscope (SEM) images of crystals observed in complexes of chlorophyll with low molecular weight proteins present in the 80 % (A) and 100 % (B) acetone-soluble fractions.

By using an SEM/energy dispersive X-ray (EDX) analyzer, Fe atoms were detected at point A (A) and S and Mo atoms were detected at point D (B).

ferences in the chemical structure of their Fe-S centers. In *E. dispar*, Fd is considered as an essential redox protein involved in energy metabolism, similar to that in *E. histolytica*, although neither species contains Mt. These findings suggest that *E. dispar* grown under axenic culture conditions may lack a sufficient quantity of some essential component of the Fe-S proteins (e.g., Fe-S centre).

The growth-promoting effect of the GPF was tested on three other strains of *E. dispar* (SAW 1734R clone AR, AS 2 IR and CYNO 16:TPC), (Koyabashi *et al.*, 2005); it was confirmed that the GPF produced the same effect on their growth (data not shown).

DISCUSSION

Previously, we designed the YIGADHA-S medium (Kobayashi *et al.*, 2005) for axenic cultivation of *E. dispar*. It contains dihydroxyacetone (DHA) that has a significant growth-promoting effect on *E. dispar*. DHA is a ketotriose and functions as a sugar source for *E. dispar*; therefore, it is considered to be directly metabolized to DHA phosphate, which is an intermediary metabolite in the Embden-Meyerhof-Parnas gly-

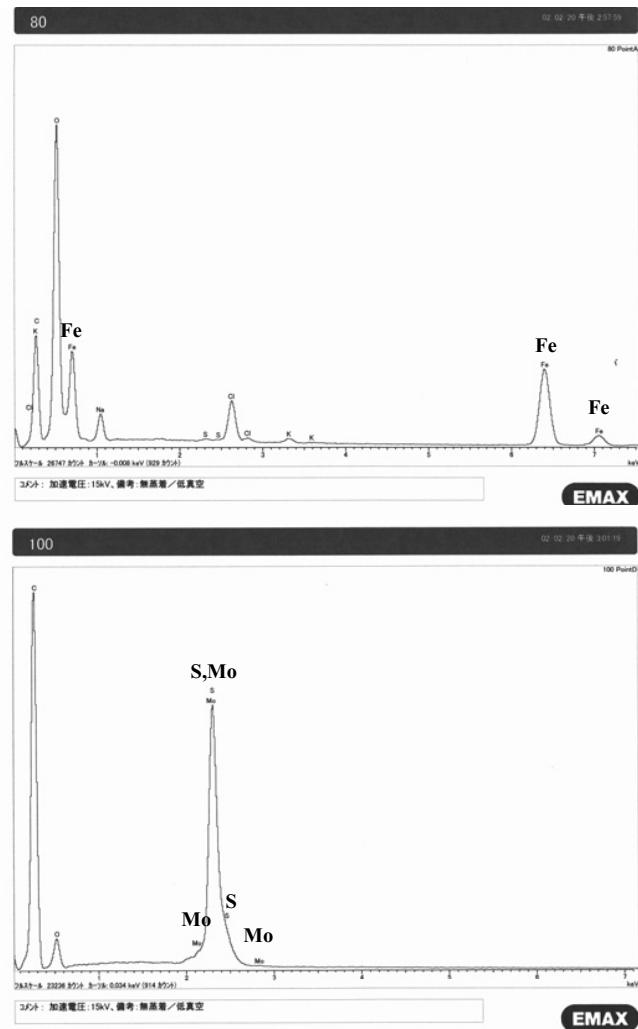


Fig. 5. – Specific peaks of the Fe atom at point A (Fig. 3) and S and Mo atoms at point D (Fig. 3) detected in the complexes of chlorophyll with low molecular weight proteins in the 80 % (A) and 100 % (B) acetone-soluble fractions by using an SEM/EDX analyzer.

colytic pathway. However, the growth of *E. dispar* was poor despite the presence of DHA.

In the present study, a GPF was detected in the autoclaved Mt, Hg and Cp fractions and in intact bacteria and was demonstrated to show a growth-promoting effect on *E. dispar*. However, the efficacy of each GPF from these organelles varied under different conditions. Prior to the present study, detection of a GPF from trophozoites of *E. histolytica* (HM-1:IMSS clone 6 strain) was attempted. Since *E. histolytica* is genetically closely related to *E. dispar*, it contains an abundance of Fe-S proteins, such as Fd, which are only stable under anaerobic conditions, and it contains mitochondrial remnant mitosomes (Leon-Avila & Tovar, 2004). However, neither the intact cells nor any extract from the cells could promote the growth of *E. dispar* AS 16 IR (data not shown). One reason for the failure to extract a GPF from *E. histolytica* trophozoites may be the fragility of

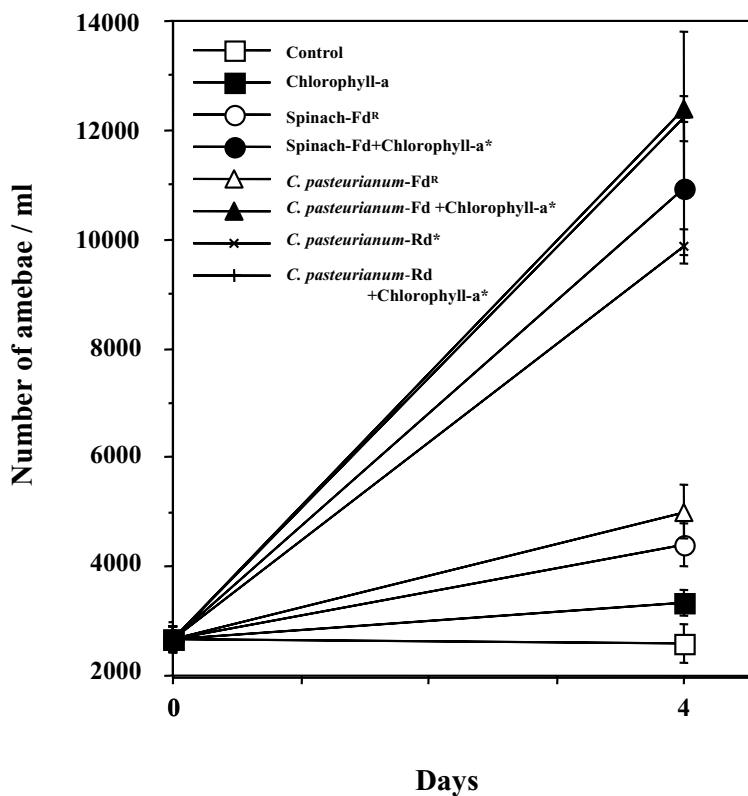


Fig. 6. - Effect of chlorophyll-a (■), 0.6 % 2ME-treated spinach ferredoxin (Fd) (○), *Clostridium pasteurianum* Fd (△) and *C. pasteurianum* rubredoxin (Rd) (×), and complexes of chlorophyll a with spinach Fd (●), *C. pasteurianum* Fd (▲) and *C. pasteurianum* Rd (→) on the growth of *E. dispar*, and control (□).

The growth kinetics of the *E. dispar* AS 16 IR strain in the YIGADHA-S medium are shown (mean numbers of amoebae in duplicate cultures are plotted). *: the mean of the growth-kinetic level was significantly higher than that of the control (* $p < 0.01$ by Dunnett's test). R: the significant difference was retained by Dunnett's test ($p < 0.05$).

the cell membrane and organelles (e.g., mitosomes) similar to that of the spinach leaf cells. The GPF was assumed to be composed of a heat-stable substance and some other common simple inorganic substances. It was feasible to isolate the GPF from acetone-soluble fractions of Cp-rich leaf cells as bands of low molecular weight proteins complexed with chlorophyll containing Fe, S and Mo atoms.

The results of this study indicate that the protein bands may be derived from Fe-S (e.g., Fd) or Mo-Fe-S proteins [e.g., molybdoferredoxin, which is a common component of nitrogenase (present in rhizobium species)]. It appears that the GPF requires to form a complex with chlorophyll because the growth-promoting effects of purified Fd from spinach, *C. pasteurianum* and *E. histolytica* strain HM-1: IMSS clone 6 (the Fd consists of 4Fe-4S centre) on *E. dispar* AS 16 IR were not remarkable (data not shown). We found that the complex of chlorophyll-a with 0.6 % 2ME-treated Fd containing a 2Fe-2S centre from spinach and that containing 4Fe-4S centre from *C. pasteurianum* have a growth-promoting effect on *E. dispar* AS 16 IR. This effect was also observed with 0.6 % 2ME-treated Rd, which has the most simple Fe-S centre (1 Fe and 4 cysteine-residues without inorganic S), without chlorophyll-a. It is not clearly understood why complex formation must occur between chlorophyll-a and the components (Fe-S centres and cysteine residues) of bacterial and plant Fd to produce a growth-promoting

effect is not clearly understood. However, it is considered that the complex of chlorophyll-a with the Fd component (MW greater than 100,000 by gel permeation chromatography) may be efficiently taken up by amoeba cells as a compact mass by a process, such as pinocytosis, and the complex may be degraded until an appropriate size of the Fe-S compound is obtained. We speculate that the Fe-S compounds are utilized as components of some essential Fe-S proteins such as amoebic Fd, which is an essential redox protein involved in oxidative decarboxylation of pyruvate to acetyl CoA in *E. dispar*; this is similar to that reported in *E. histolytica* (McLaughlin & Aley, 1985). It is reported that the amitochondrial protozoan parasite *E. histolytica*, which is closely related to *E. dispar*, contains numerous fermentation enzymes that are similar to bacterial enzymes, including the pyruvate Fd oxidoreductase, Fd and alcohol dehydrogenase E, derived from concomitant enteric bacterial DNA by horizontal transfer (Rosenthal *et al.*, 1997).

We succeeded in developing a more stable axenic cultivation system by adding the complex of chlorophyll with a component of Fe-S protein (e.g., Fd from spiderwort) treated with 2ME to the YIGADHA-S medium. We achieved an improved axenic cultivation system for *E. dispar* by using cellular components of protozoan, mammalian and plant cells as well as intact bacteria. However, we recognize that it is difficult to develop an axenic cultivation system for *E. dispar* that is at par

with the well-established cultivation system of *E. histolytica* for use in biological studies such as biochemical and immunological studies, etc.

ACKNOWLEDGEMENTS

The authors thank the staff of Nihonbashi Laboratory, Hitachi High-Technologies Co., Tokyo, Japan for their cooperation in the analysis using the SEM-EDX III Type N/H. This work was supported by a Keio Gijuku Fukuzawa Memorial Fund for the Advancement of Education and Research and a Health Sciences Research Grant-in-Aid for Emerging and Re-emerging Infectious Diseases.

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Reçu le 14 juin 2005

Accepté le 4 novembre 2005