

# HELMINTH COMMUNITIES OF TWO GREEN FROGS (*RANA PEREZI* AND *RANA SAHARICA*) FROM BOTH SHORES OF THE ALBORAN SEA

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## Summary:

The helminth communities of two populations of green frogs from both shores of the Alborán Sea (Western Mediterranean) were studied. Of the 79 frogs examined for helminths, 39 individuals of the species *Rana saharica* were collected from Bab-Taza (Morocco), and 40 of the species *Rana perezi* were collected from the Natural Park of the Sierra de Grazalema (Spain). Although the species richness of helminths was identical in the two sampled areas, the differences observed in the structure of the helminth infracommunities were quite important. Statistically, significant differences were found between the species richness and the diversity of the infracommunities of *R. perezi* female population and the other three studied statistical populations. The helminth component communities of these two green frogs can be considered as depauperate, although their infracommunities present interactive features.

**KEY WORDS :** helminth communities, helminth infracommunities, *Rana perezi*, *Rana saharica*, species richness, diversity.

**Résumé :** COMMUNAUTÉS D'HELMINTHES DE DEUX POPULATIONS DE GRENOUILLES VERTES (*RANA PEREZI* ET *RANA SAHARICA*) DES DEUX RIVAGES DE LA MER D'ALBORÁN

Les communautés d'helminthes de deux populations de grenouilles vertes des rivages de la mer d'Alborán (méditerranéen occidental) ont été étudiées. Des 79 grenouilles examinées pour étudier les helminthes, 39 exemplaires de *Rana saharica* ont été capturés à Bab-Taza (Maroc), et 40 de l'espèce *Rana perezi* dans le Parc Naturel Sierra de Grazalema (Espagne). Bien que la richesse des espèces d'helminthes soient identique dans les deux secteurs d'échantillonnage, les différences observées dans la structure des infracommunautés d'helminthes sont assez importantes. Statistiquement, des différences significatives ont été trouvées entre la richesse en espèces et la diversité des infracommunautés chez les femelles de *R. perezi* et les trois autres populations statistiques étudiées. Les communautés helminthiques de ces deux grenouilles vertes peuvent être considérées comme pauvres, bien que leurs infracommunautés présentent des caractéristiques interactives.

**MOTS CLÉS :** communauté d'helminthes, infracommunauté d'helminthes, *Rana perezi*, *Rana saharica*, richesse spécifique, diversité.

## INTRODUCTION

Most of the studies related to parasite fauna of European and African amphibians deal mainly with faunistic aspects (Baker, 1981; Murai *et al.*, 1983; Lluch *et al.*, 1986a, 1986b; Kok & Seaman, 1987; Moravec *et al.*, 1987; Kok, 1989; Malashichev, 2000; Galli *et al.*, 2001; Aisien *et al.*, 2003; du Preez *et al.*, 2003).

The revision carried out by Aho (1990) on the helminth community structure of amphibians indicates that the corresponding communities can be highly variable, although they are habitually depauperate and noninteractive in structure.

Various authors (Muzzall, 1991; Goldberg *et al.*, 1995; McAlpine, 1997; Muzzall *et al.*, 2001; Bolek & Coggins, 2003) consider that amphibian populations provide excellent systems to study ecological concepts related to helminth communities. Therefore, the study of hel-

minth communities in two species of green frogs, *Rana perezi* Seoane, 1885 and *Rana saharica* Bolu-lenger, 1913, has been carried out. The studied samples were collected from two sites of the Betico-Rifeña Chain, one on each side of the Alborán Sea. The *R. perezi* specimens were taken from the Natural Park of the Sierra de Grazalema (Spain), and the specimens of *R. saharica* were taken from Bab-Taza (Morocco). Since the studied ranids are vicariant (Pleguezuelos *et al.*, 2002) and similar in corporal size, physiology and trophic behaviour (Bons & Geniez, 1996; Esteban *et al.*, 1999), and there are no dietary differences between males and females of both species (Garcia-Paris, 1985; Lizana *et al.*, 1990; Meddeb & Cheniti, 1998), the observed helminth communities would provide an important information dealing with biotic and abiotic factors which regulate the structure of these communities in relation to the habitat conditions. In addition, as both sampling sites are located in the same mountainous chain and both are directly exposed to the Mediterranean influence, it should be expected that the helminth communities of the host populations were similar and also, as it has been previously commented (Aho, 1990), depauperate and noninteractive in structure.

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## MATERIALS AND METHODS

40 individuals of *Rana perezi* (18 females and 22 males) were collected in June 1993 from the portion of Cádiz of the Sierra de Grazalema Natural Park (Spain), and 39 specimens of *Rana sabarica* (18 females and 21 males) were collected in September 1997 from Bab-Taza, in the south of Chefchaouen (Western Rif, Morocco). Euthanasia of hosts was performed by an overdose of inhalant anesthetic (ethyl acetate) in accordance with the Canadian Council on Animal Care (1993) guidelines. Data relative to host sex were taken after gonadal inspection and the measures corresponding to the total length (mm) of each frog were recorded. Eventual differences in body length between males and females of the two host species were tested using a one-way analysis of variance (ANOVA). Necropsies of complete specimens were performed and all helminths were counted and identified. Levels of parasitic load and species richness in the four statistical host populations were compared using nonparametric tests, the Kruskal-Wallis test and the Mann-Whitney U test. The parasites were processed following conventional techniques. Trematodes and acanthocephalans were fixed in aqueous Bouin and conserved in 70° ethanol, stained with Carmine alum, dehydrated and mounted with Canada balsam. Nematodes were fixed and conserved in 70° ethanol, and were studied using Amann lactophenol wet mounts. All the helminths found, as well as all host carcasses have been deposited in the collection of the Departamento de Zoología of the University of Valencia (Spain).

The terms prevalence, mean intensity and mean abundance relative to the parasite populations have been used as defined by Bush *et al.*, (1997). A helminth species was considered as common if it occurred in 50 % or more of hosts (Aho, 1990; Muzzall *et al.*, 2001). The mean species richness is the mean number of helminth species per examined frog. The community species richness is the number of helminth species found in an entire host population, and the community abundance is the total number of individual helminths found in an entire host population. Diversity refers to the composition of a sample in terms of helminth species number found in such sample, and it includes a factor that evaluates the evenness in the relative distribution of each helminth species. Evenness is a measurement of the disparity in the number of individuals found for each parasite species. Some authors, such as Poulin (1998), have pointed out some defects of the diversity indexes, indicating that they can be misleading when large parasites coexist in the same host with others of smaller size. In this study we will consider these indexes as simple descriptors of properties of the studied infracommunities, since they can be useful to

establish comparisons with other studies and since the parasitic fauna of the green frogs does not usually include large cestodes that mask the results. Values of Brillouin's index (Pielou, 1975) were calculated for measurements of helminth infracommunity diversity and evenness, by means of the DIVERS programme (Krebs, 1989) and the subsequent common logarithms transformation. For component communities, Shannon-Wiener's index, because it is less biased toward dominant species than other indices (Marcogliese, 2002), and Berger-Parker index were calculated for diversity and dominance analysis respectively, using the programme of Pérez-López & Solá-Fernández (1993).

A correlation analysis was performed between the size of the individuals from the four statistical populations and the diversity, the evenness, the species richness and the abundance of the corresponding helminth infracommunities, using Spearman's coefficient, due to the low number of values obtained (Ludwig & Reynolds, 1988).

An eventual influence of the host sex on parasite absolute frequencies, as well as a possible establishment of associations between pairs of helminth species were analysed using G-test, with Wilian's correction formula. Likewise, Mann-Whitney U test was used to determine whether there was an influence of the host sex on the abundance of parasites.

## RESULTS

The overall data relative to prevalence, mean intensity and mean abundance of the parasites detected in both sexes of *Rana perezi* and *Rana sabarica* are summarised in Table I. No helminth species could be considered common in the female hosts of *R. perezi*, and only *Rhabdias bufonis* was common in the males of this host species. In contrast, *Opisthodiscus nigrivasis*, *Cosmocerca ornata* and *Spiroxys contortus* were common parasite species in the females of *R. sabarica*, and *R. bufonis* and *C. ornata* in their respective males.

In addition, the comparative study of the helminth communities of the two host species allowed to state that, although the abundance of helminths did not show statistically significant differences (Mann-Whitney U-test,  $P = 0.566$ ), the total number of parasites was much higher in *R. perezi* (1029) than in *R. sabarica* (538). Table II presents the results provided by the analysis of helminth communities and helminth infracommunities in which host species and host sex were taken into account. Considering a mean number of parasite species, females of *R. perezi* were significantly less infected than the males of the two studied amphibians and than the females of *R. sabarica* (Kruskal-Wallis test,  $P = 0.016$

Frog species Helminth species	Location in host	Mean intensity $\pm$ SD (range)	Prevalence	Mean abundance $\pm$ SD
<i>Rana perezi</i> females				
<i>D. subclavatus</i>	L. in	2.50 $\pm$ 2.12 (1-4)	2/18 (11.1 %)	0.28 $\pm$ 0.96
<i>O. nigrivasis</i>	L. in	2.00 $\pm$ 0.00 (2)	1/18 (5.5 %)	0.11 $\pm$ 0.47
<i>G. microoovata</i>	B	1.67 $\pm$ 1.15 (1-3)	3/18 (16.6 %)	0.28 $\pm$ 0.75
<i>L. nigrovenosus</i>	Sk	7.00 $\pm$ 0.00 (7)	1/18 (5.5 %)	0.39 $\pm$ 1.65
<i>C. europaeus</i>	S. in	3.50 $\pm$ 2.12 (2-5)	2/18 (11.1 %)	0.39 $\pm$ 1.24
<i>P. stromi</i>	S. in	1.00 $\pm$ 0.00 (1)	1/18 (5.5 %)	0.06 $\pm$ 0.24
<i>S. tacapense</i>	S. in	1.00 $\pm$ 0.00 (1)	1/18 (5.5 %)	0.06 $\pm$ 0.24
<i>R. bufonis</i>	L	4.14 $\pm$ 3.58 (1-11)	7/18 (38.8 %)	1.61 $\pm$ 2.97
<i>S. numidica</i>	S. in	4.80 $\pm$ 3.77 (1-9)	5/18 (27.7 %)	1.33 $\pm$ 2.87
<i>C. ornata</i>	S. in	18.43 $\pm$ 28.17 (1-78)	7/18 (38.8 %)	7.17 $\pm$ 19.12
<i>A. macintoshii</i>	S. in	12.00 $\pm$ 7.97 (1-22)	5/18 (27.7 %)	3.33 $\pm$ 6.75
<i>I. neglecta</i>	Mc	1.50 $\pm$ 0.71 (1-2)	2/18 (11.1 %)	0.17 $\pm$ 0.51
<i>Rana perezi</i> males				
<i>D. subclavatus</i>	L. in	2.20 $\pm$ 1.64 (1-5)	5/22 (22.7 %)	0.50 $\pm$ 1.19
<i>O. nigrivasis</i>	L. in	2.20 $\pm$ 0.45 (2-3)	5/22 (22.7 %)	0.50 $\pm$ 0.96
<i>H. kessleri</i>	St	1.00 $\pm$ 0.00 (1)	1/22 (4.5 %)	0.05 $\pm$ 0.21
<i>G. microoovata</i>	B	7.86 $\pm$ 11.74 (1-34)	7/22 (31.8 %)	2.50 $\pm$ 7.31
<i>G. vitelliloba</i>	B	1.00 $\pm$ 0.00 (1)	2/22 (9.1 %)	0.09 $\pm$ 0.29
<i>C. europaeus</i>	S. in	8.22 $\pm$ 8.26 (2-25)	9/22 (40.9 %)	3.36 $\pm$ 6.57
<i>P. stromi</i>	S. in	64.00 $\pm$ 75.94 (1-171)	5/22 (22.7 %)	14.55 $\pm$ 43.04
<i>S. tacapense</i>	S. in	6.10 $\pm$ 7.05 (1-22)	10/22 (45.5 %)	2.77 $\pm$ 5.56
<i>S. joyeuxi</i>	Mc	57.00 $\pm$ 0.00 (57)	1/22 (4.5 %)	2.59 $\pm$ 12.15
<i>R. bufonis</i>	L	6.08 $\pm$ 8.33 (1-27)	13/22 (59.1 %)	3.59 $\pm$ 7.00
<i>S. numidica</i>	S. in	1.50 $\pm$ 0.58 (1-2)	4/22 (18.2 %)	0.27 $\pm$ 0.63
<i>C. ornata</i>	S. in	5.11 $\pm$ 4.14 (1-14)	9/22 (40.9 %)	2.09 $\pm$ 3.62
<i>A. macintoshii</i>	S. in	3.13 $\pm$ 2.64 (1-8)	8/22 (36.4 %)	1.14 $\pm$ 2.17
<i>I. neglecta</i>	Mc	1.75 $\pm$ 0.96 (1-3)	4/22 (18.2 %)	0.32 $\pm$ 0.78
<i>A. falcatus</i>	S. in	1.00 $\pm$ 0.00 (1)	1/22 (4.5 %)	0.05 $\pm$ 0.21
<i>Rana sabarica</i> females				
<i>D. subclavatus</i>	L. in	14.00 $\pm$ 19.92 (2-37)	3/18 (16.7 %)	2.33 $\pm$ 8.69
<i>O. nigrivasis</i>	L. in	2.29 $\pm$ 0.83 (1-3)	14/18 (77.8 %)	1.78 $\pm$ 1.22
<i>G. microoovata</i>	B	4.33 $\pm$ 4.16 (1-9)	3/18 (16.7 %)	0.72 $\pm$ 2.19
<i>L. nigrovenosus</i>	Sk	1.75 $\pm$ 0.96 (1-3)	4/18 (22.2 %)	0.39 $\pm$ 0.85
<i>P. chamaeleonis</i>	S. in	1.50 $\pm$ 0.71 (1-2)	2/18 (11.1 %)	0.17 $\pm$ 0.51
<i>S. joyeuxi</i>	Mc	3.00 $\pm$ 0.00 (3)	1/18 (5.6 %)	0.17 $\pm$ 0.71
<i>R. bufonis</i>	L	5.00 $\pm$ 5.29 (1-16)	7/18 (38.9 %)	1.94 $\pm$ 4.02
<i>S. numidica</i>	S. in	1.83 $\pm$ 1.17 (1-4)	6/18 (33.3 %)	0.61 $\pm$ 1.09
<i>C. ornata</i>	S. in	2.89 $\pm$ 1.54 (1-5)	9/18 (50.0 %)	1.44 $\pm$ 1.82
<i>A. macintoshii</i>	S. in	2.86 $\pm$ 1.86 (2-7)	7/18 (38.9 %)	1.11 $\pm$ 1.81
<i>Paracamallanus</i> sp.	St	1.00 $\pm$ 0.00 (1)	1/18 (5.6 %)	0.06 $\pm$ 0.24
<i>S. contortus</i>	Mc	6.78 $\pm$ 4.21 (1-12)	9/18 (50.0 %)	3.39 $\pm$ 4.53
Physalopteridae gen. sp.	S. in; L. in	2.00 $\pm$ 1.41 (1-3)	2/18 (11.1 %)	0.22 $\pm$ 0.73
<i>Rana sabarica</i> males				
<i>D. subclavatus</i>	L. in	2.25 $\pm$ 1.50 (1-4)	4/21 (19.0 %)	0.43 $\pm$ 1.08
<i>O. nigrivasis</i>	L. in	1.83 $\pm$ 0.98 (1-3)	6/21 (28.6 %)	0.52 $\pm$ 0.98
<i>G. microoovata</i>	B	2.00 $\pm$ 1.41 (1-5)	8/21 (38.1 %)	0.76 $\pm$ 1.30
<i>H. variegatus</i>	L	1.00 $\pm$ 0.00 (1)	1/21 (4.8 %)	0.05 $\pm$ 0.22
<i>S. joyeuxi</i>	Mc	8.50 $\pm$ 2.12 (7-10)	2/21 (9.5 %)	0.81 $\pm$ 2.60
<i>R. bufonis</i>	L	5.18 $\pm$ 5.21 (1-15)	11/21 (52.4 %)	2.71 $\pm$ 4.54
<i>S. numidica</i>	S. in	2.29 $\pm$ 1.25 (1-4)	7/21 (33.3 %)	0.76 $\pm$ 1.30
<i>C. ornata</i>	S. in	3.92 $\pm$ 2.31 (1-10)	12/21 (57.1 %)	2.24 $\pm$ 2.62
<i>A. macintoshii</i>	S. in	5.40 $\pm$ 4.72 (2-13)	5/21 (23.8 %)	1.29 $\pm$ 3.16
<i>S. contortus</i>	Mc	8.44 $\pm$ 14.30 (1-46)	9/21 (42.9 %)	3.62 $\pm$ 10.01
Physalopteridae gen. sp.	S. in; L. in	1.00 $\pm$ 0.00 (1)	1/21 (4.8 %)	0.05 $\pm$ 0.22
<i>Pseudabbreviata</i> sp.	S. in	1.00 $\pm$ 0.00 (1)	1/21 (4.8 %)	0.05 $\pm$ 0.22
<i>Agamospirura</i> sp.	Mc	1.00 $\pm$ 0.00 (1)	1/21 (4.8 %)	0.05 $\pm$ 0.22

Table I. – Location, mean intensity, prevalence and mean abundance of helminth parasites recovered from males and females of *Rana perezi* and *Rana sabarica* (L. in = large intestine; B = bladder; Sk = skin; S. in = small intestine; L = lungs; Mc = muscles; St = stomach).

Host populations	<i>R. perezii</i> (n = 40)	<i>R. perezii</i> females (n = 18)	<i>R. perezii</i> males (n = 22)	<i>R. sabarica</i> (n = 39)	<i>R. sabarica</i> females (n = 18)	<i>R. sabarica</i> males (n = 21)
R	16	12	15	16	13	13
A	1029	273	756	538	258	280
H'	2.13	1.60	1.94	2.17	2.17	2.02
E	0.77	0.64	0.72	0.78	0.85	0.79
d	0.31	0.47	0.42	0.26	0.24	0.27
Dominant species	<i>P. stromi</i>	<i>C. ornata</i>	<i>P. stromi</i>	<i>S. contortus</i>	<i>S. contortus</i>	<i>S. contortus</i>
Mean species richness ± SD (range)	3.03 ± 2.08 (0-7)	2.06 ± 1.86 (0-6)	3.82 ± 1.94 (0-7)	3.49 ± 1.55 (0-6)	3.78 ± 1.52 (1-6)	3.24 ± 1.58 (0-6)
Mean helminth abundance ± SD (range)	25.73 ± 36.85 (0-175)	15.17 ± 20.32 (0-78)	34.36 ± 44.88 (0-175)	13.79 ± 12.16 (0-58)	14.33 ± 12.73 (1-58)	13.33 ± 11.94 (0-53)
Mean Brillouin diversity (ln) ± SD (range)	0.57 ± 0.44 (0.00-0.52)	0.38 ± 0.40 (0.00-1.19)	0.73 ± 0.42 (0.00-1.52)	0.72 ± 0.36 (0.00-1.26)	0.80 ± 0.34 (0.00-1.26)	0.66 ± 0.37 (0.00-1.17)
Mean helminth evenness ± SD (range)	0.58 ± 0.40 (0.00-1.00)	0.43 ± 0.43 (0.00-1.00)	0.70 ± 0.34 (0.00-1.00)	0.74 ± 0.32 (0.00-1.00)	0.81 ± 0.24 (0.00-1.00)	0.68 ± 0.37 (0.00-1.00)
Proportion of sample with 1 or 0 helminth species	0.25	0.45	0.09	0.13	0.06	0.19

Table II. – Diversity characteristics of biological and statistical populations studied. R = species richness; A = helminth abundance; H' = diversity (Shannon-Wiener index); E = evenness; d = dominance (Berger-Parker index).

for the four statistical populations; Mann-Whitney U-test,  $P = 0.008$  for males and females of *R. perezii*,  $P = 0.007$  for the females of both anurous,  $P = 0.034$  for the females of *R. perezii* and the males of *R. sabarica*). The low number of helminth species that infest the females of *R. perezii* determines that the diversity of their helminth infracommunities is statistically inferior to that of the infracommunities corresponding to both, males of the same species (ANOVA  $P = 0.010$ ) and statistical populations of females and males of *R. sabarica* (ANOVA,  $P = 0.002$  and  $P = 0.032$ , respectively). Equally, the evenness value for the infracommunities of the statistical population of *R. perezii* females was significantly lower than both, the one for the infracommunities of males of the same species, and the one corresponding to *R. sabarica* females (ANOVA,  $P = 0.028$  and  $P = 0.002$ , respectively). Nevertheless, this evenness did not significantly vary from the one of the statistical population of *R. sabarica* males (ANOVA,  $P = 0.052$ ).

The differences observed among the four helminth infracommunities can not be attributable to host size, since the comparison of the total length of hosts showed no significant differences among the four studied statistical populations (ANOVA,  $P = 0.151$ ).

The analysis of host sex in relation to parasite absolute frequencies, showed that the occurrence of the helminths *O. nigrivasis* and *Leptophallus nigrovenosus* was significantly more frequent in females of *R. sabarica* than in males of this species (G-test,  $P < 0.01$  and

$P < 0.02$ , respectively), and the respective abundances were also higher in females of *R. sabarica* than in males of this species (Mann-Whitney U-test,  $P = 0.002$  and  $P = 0.025$ , respectively). Besides, *Sonsinotrema tacapense* and *Cephalogonimus europaeus* occurred more frequently in males of *R. perezii* than in females of this ranid (G test,  $P < 0.01$  and  $P < 0.05$ , respectively) and, as in the previous case, the respective abundances were also higher in males of *R. perezii* than in females of this species (Mann-Whitney U-test,  $P = 0.004$  and  $P = 0.033$ , respectively).

In the statistical population of *R. perezii* females, the species richness (Spearman's correlation  $r_s = 0.637$ ,  $P = 0.005$ ) and the abundance of the helminth infracommunities (Spearman's correlation  $r_s = 0.72$ ,  $P = 0.001$ ) were positively and significantly correlated with the host size. This correlation was not found in the other three statistical populations of the anurans.

Ten positive and one negative associations between pairs of helminth species were found in *R. perezii*, and one positive and four negative associations were found in *R. sabarica* (Table III).

## DISCUSSION

The comparative study of the helminth infracommunities in the two amphibian species showed that the mean abundance of helminths was

	Sign	G <sub>adj,1df</sub>	P
<b><i>Rana perezi</i> associations (n = 40)</b>			
<i>O. nigrivasis</i> - <i>C. europaeus</i>	+	4.34	0.05 > P > 0.02
<i>O. nigrivasis</i> - <i>C. ornata</i>	+	5.16	0.05 > P > 0.02
<i>G. microovata</i> - <i>S. tacapense</i>	+	6.12	0.02 > P > 0.01
<i>C. europaeus</i> - <i>S. tacapense</i>	+	4.90	0.05 > P > 0.02
<i>C. europaeus</i> - <i>R. bufonis</i>	+	6.21	0.02 > P > 0.01
<i>C. europaeus</i> - <i>S. numidica</i>	+	3.92	0.05 > P > 0.02
<i>P. stromi</i> - <i>A. macintoshii</i>	+	7.21	0.01 > P > 0.001
<i>R. bufonis</i> - <i>C. ornata</i>	+	6.64	0.01 > P > 0.001
<i>R. bufonis</i> - <i>A. macintoshii</i>	+	5.56	0.02 > P > 0.01
<i>C. ornata</i> - <i>I. neglecta</i>	+	5.16	0.05 > P > 0.02
<i>O. nigrivasis</i> - <i>A. macintoshii</i>	-	4.73	0.05 > P > 0.02
<b><i>Rana sabarica</i> associations (n = 39)</b>			
<i>A. macintoshii</i> - Physalopteridae g. sp.	+	5.20	0.02 > P > 0.01
<i>O. nigrivasis</i> - <i>D. subclavatus</i>	-	4.72	0.05 > P > 0.02
<i>Szidatia joyeuxi</i> - <i>C. ornata</i>	-	4.22	0.05 > P > 0.02
<i>S. numidica</i> - <i>S. contortus</i>	-	4.17	0.05 > P > 0.02
<i>C. ornata</i> - <i>A. macintoshii</i>	-	9.74	0.01 > P > 0.001

Table III. – Associations between pairs of helminth species in *Rana perezi* and *Rana sabarica*.

higher in *R. perezi* than in *R. sabarica*. Females of *R. perezi* presented the particularity of being different from the other three statistical populations in the helminth infracommunities. In fact, species richness and diversity were significantly smaller in the helminth infracommunities of *R. perezi* females. Moreover, the proportions of parasite-free hosts or hosts infected with only one parasite species were significantly higher in these females, which were also different from the other populations in the absence of common parasite species.

Species richness and abundance of the helminth communities (Table II) were lower in the statistical population of *R. perezi* females than in males of this host species. Diversity and evenness values were also lower in *R. perezi* females than in all the other three statistical populations, and the parasite species with the highest value of dominance in females of *R. perezi* was *Cosmocerca ornata*. In addition, significant correlations of host length/species richness and host length/abundance were exclusively established in the females of *R. perezi*.

Since some of the previously alluded differences, as those referring to the mean species richness and the diversity of the infracommunities, were statistically significant, it has been attempted to establish the causes of such dissimilarities. According to Kennedy *et al.* (1986), one of the major factors which contribute to the diversity of helminths is a complex diet of the host that facilitates the ingestion of different intermediate hosts. Among the helminths found, only three species were monoxenous and the cercariae of some trematodes penetrated in the definitive host through the skin, but other helminths invaded their hosts by means of their feeding habits. Although in other studies dealing with the diet of green frogs (Garcia-Paris, 1985; Lizana

*et al.*, 1990; Meddeb & Cheniti, 1998), no dietary differences between male and female anurans were mentioned, it is obvious that larger host individuals must ingest greater amount of prey items. However, the analysis of host size did not highlight any significant difference between *R. perezi* females and the individuals of the other three statistical populations. Therefore, the factor host size can not be used to explain the dissimilarities detected in the helminthofauna of *R. perezi* females. In spite of this, females of *R. sabarica* (mean snout-vent length = 53.44 + 8.43 mm) were slightly larger than males (mean snout-vent length = 51.55 + 9.47 mm). This could justify the slightly bigger, but not statistically significant, parasitic load observed in the females of this species.

Various authors (Poulin, 1996; Zuk & McKean, 1996; Schalk & Forbes, 1997; Klein, 2000a) affirm that the males of many animal species are more susceptible than their respective females to parasite infections due to hormonal or biological factors. Lees (1962) pointed out that the greatest difference in the levels of parasitization in males and females of *Rana temporaria* occurred when a high level of sex hormone in the blood was observed. However, Plasota (1969) affirms that the differences between males and females of *Rana esculenta* and *Rana terrestris* is not the result of hormonal differences but mostly depends on the biological differences between the two sexes.

Males and females of *R. sabarica* were collected in September, far from the reproductive season, when the hormonal levels were low. Besides, specimens of *R. perezi* were collected in June, during the reproductive season, when the hormonal levels were presumably the highest. Therefore, the hormonal factor and the estrogen protection against determined parasites could be the cause of the observed dissimilarities. This explanation is

consistent with the fact that there were very slight differences between the helminth infracommunities of males and females of *R. sabarica*, with an expected low level of estrogen in the females. These differences were evident and statistically significant between the helminth infracommunities of males and females of *R. perezi* whose females would contain high levels of estrogens. It is also necessary to take into consideration that exposure to environmental and social stressors can suppress immune function and increase susceptibility to disease (Klein, 2000b). Several studies in Amphibia illustrate that courtship displays are costly from the metabolism point of view, being able to increase male susceptibility to infection (Taigen & Wells, 1985; Pfennig & Tinsley, 2002). This factor could have contributed to increase the parasitic load of *R. perezi* males.

The results obtained by Muzzall (1991) and McAlpine (1997), in relation to the analysis of the helminth communities of different holarctic frogs, are consistent with those provided hereby on the helminth infracommunities of green frogs, although the diversity and the evenness values are slightly higher in the present study. Nevertheless, the species richness, the diversity and the abundance values of the helminth infracommunities were very low in these studied green frogs compared to the values found in the helminth infracommunities of *Rana clamitans* from Michigan, USA (Muzzall *et al.*, 2001). These authors set that the helminth communities of *R. clamitans* were depauperate and noninteractive.

The present study shows that the helminth communities of the examined frogs were depauperate, but they should be considered as interactive, since there were (Table III) many positive and negative associations built up between pairs of helminths of both *R. perezi* and *R. sabarica* populations. Lotz & Font (1994) and Poulin (1997) indicated that a high proportion of rare parasite species (prevalence less than 10 %) in the component community, can lead to an excess of negative associations, and that a high proportion of common species (prevalence greater than 90 %), can produce an excess of positive associations. Among the obtained results, the studied component communities do not show unusual proportions of rare or common species, for what they can represent interactive communities. Positive interactions in *R. perezi* can be given because the presence of a parasite species, can facilitate the infection by other species and negative interactions in *R. sabarica* can have their origin in factors such as competitive exclusion.

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cript. The authors declare that the experiments comply with the current laws of the country in which they were performed.

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