ASSESSMENT OF IMMUNOCHROMATOGRAPHIC TEST FOR RAPID LYMPHATIC FILARIASIS DIAGNOSIS

NGUYEN N.L.*, PLICHART C.** & ESTERRE P.**

Summary:

Two rapid immunodiagnostic tests (ICT Filariasis test®), developed for the quick diagnosis of Wuchereria bancrofti infection, have been validated in laboratory and field situation. The aim of this study was to assess the performance and usefulness of this antigen capture assay as a diagnostic method in three foci of lymphatic filariasis, located in the South Pacific (Society archipelago, French Polynesia), with different levels of endemicity. A sample of 1,595 patients was tested with this assay in parallel with a reference Og4C3 antigen capture assay and microfilariae detection. A second-generation ICT test, available for whole blood analysis, was also tested in parallel with the first generation test, developed for serum analysis, on a sample of 50 reference cases. The correspondence between the results obtained with the two rapid tests was excellent, without any influence of rheumatoid factors, but the sensitivity was in both cases slightly inferior to the ELISA reference test. This seems particularly true in epidemiological situation where a high proportion of microfilaraemic, adult worm carriers are observed.

KEY WORDS: lymphatic filariasis, Wuchereria bancrofti, rapid antigen capture assay, ICT Filariasis test, French Polynesia.

Résumé : Validation de tests immunochromatographiques pour le diagnostic rapide de la filariose lymphatique

Deux tests d’immunodiagnostic rapide de la filariose à Wuchereria bancrofti (kits australiens ICT Filariose®) ont été étudiés sur le terrain et en laboratoire. Le but de cette étude était de vérifier les performances et l’utilité réelle de ces tests de capture d’antigène dans la filariose lymphatique dans trois foyers d’endémicité différente, situés dans l’archipel de la Société (Polynésie Française). Un échantillon de 1 593 patients a donc été analysé en parallèle avec le test ICT, la technique ELISA de référence (capture de l’antigène circulant Og4C3) et la détection de microfilaires sanguines (par filtration sur membrane Nucléopore®). Une seconde génération de test, validée pour l’utilisation de sang total, a été testée en parallèle sur une cinquantaine de sérum de référence, par rapport au test de première génération, validé sur le sérum uniquement. La correspondance entre les résultats obtenus par les deux tests rapides est excellente mais la sensibilité de cette technique immunochromatographique est toujours inférieure à celle de l’ELISA de référence. Contrairement à un test équivalent de diagnostic rapide du paludisme (ICT Malaria Pf®), il ne semble pas y avoir de faux positifs dus à la présence de facteurs rhumatoïdes. La performance inférieure par rapport à l’ELISA semble accentuée dans les situations épidémiologiques où l’on observe une forte proportion d’individus microfilariémiques mais porteurs de vers adultes, comme dans certaines îles du Pacifique soumises à un contrôle de masse depuis de nombreuses années.

KEY WORDS : filariose lymphatique, Wuchereria bancrofti, test rapide d’immunocapture, test “ICT Filariose”, Polynésie Française.

MATERIAL AND METHODS

Serum samples

We used 399 reference sera of inhabitants from Tahaa island, collected during a recent survey organized after four years of annual bitherapy with diethylcarbamazine (DEC) and ivermectin (IVM) (see Moulia-Pelat et al., 1995), and 200 reference sera from Opoa island, after six years of semi-annual chemotherapy with IVM (Nguyen et al., unpublished data). In addition, 996 reference sera, taken...
from all inhabitants of Maupiti island where filariasis endemic is at very low level, were used. At the 1997 end-point, the epidemiological situation in Tabaa, Opoa and Maupiti islands was quite different (see Table II).

METHODS

Following the manufacturer’s recommendations, analysis was performed on all samples for the antigenemia with a commercial (TropBio, JCU Tropical Biotech. Ltd., Townsville, Australia) ELISA assay using an IgM (called Og4C3) monoclonal antibody (Chanteau et al., 1994, Simonsen et al., 1996), and with the rapid immunochromatographic test (ICT Filarisis®, ICT Diagnostics Ltd., Brookvale, Australia) developed for serum analysis (first generation tests: Freedman et al., 1997; Weil et al., 1997) and using a monoclonal antibody called AD12.1 (Weil et al., 1987). Subsequently, on a sub-sample of 50 randomly selected sera, a second generation test (ICT Filarisis Whole Blood Test®) was performed in parallel with the first generation test (ICT Filarisis®). All samples were marked in code and tested without knowledge of previous test results. Ten rheumatoid factor positive sera, as determined by direct haemagglutination (Rhumalatex® kit, Fumouze lab., Levallois, France) were tested with the ICT test in order to check for the possibility of false positivity due to autoantibody formation (Grobusch et al., 1999). All discordant results between the quick test and the ELISA assay were checked by a second round of ICT testing.

STATISTICAL ANALYSIS

The variation between the card-test and the ELISA assay was determined by calculating Kappa (κ) values with 95% confidence intervals. The κ values between 0.60 and 0.80 were considered as good, and κ > 0.81 as very good agreement beyond chance (Buhrer et al., 1998). The McNemar’s chi-square test, based on the number of discordant pairs, was used to test for differences in the results of the standard (ELISA) and rapid (ICT) tests, and between the first- and second-generations ICT tests, according to the different groups. The sensitivity and specificity of the ICT tests were determined using the Og4C3 ELISA as the reference test.

RESULTS

On the basis of the microfilaremia only (Nucleopore membrane filtration technique of 1 ml of venous blood), the sera can be classified in two groups: amicrofilaremic and microfilaremic patients. If we also consider the results of the ELISA assay detecting the circulating Og4C3 antigen, the sera can be classified into three groups (Nicolas, 1997): microfilaremic patients, positive with both tests; amicrofilaremic “adult worm carrier” patients, with only a positive antigenemia; and negative controls. On the basis of epidemiological data, the sera were also classified following the degree of endemicity of the corresponding island.

The results of the initial analysis using the whole sample (1595 sera) are shown in Table I. A quite good degree of global agreement (92.4 %) was observed between the ELISA and the card-test but a significant difference was observed between these two tests (McNemar Chi² = 49, p < 0.0001). The agreement beyond chance was clas-

<table>
<thead>
<tr>
<th>Og4C3 ELISA</th>
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<tbody>
<tr>
<td>Positive</td>
<td>104</td>
<td>22</td>
<td>126</td>
</tr>
<tr>
<td>Negative</td>
<td>99</td>
<td>1,370</td>
<td>1,469</td>
</tr>
<tr>
<td>Total</td>
<td>203</td>
<td>1,392</td>
<td>1,595</td>
</tr>
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The agreement was 92.4 % with a kappa value of 0.60 (SE = 0.05, p < 0.0001).

Table I. Comparison between the ELISA and the ICT test results for the detection of circulating antigen of Wuchereria bancrofti.

<table>
<thead>
<tr>
<th>Microfilar prevalence (%)</th>
<th>Og4C3 antigen</th>
<th>ICT card-test results</th>
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<tr>
<td></td>
<td>Mean levela</td>
<td>+ ve (%) c</td>
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<tr>
<td>Hyperendemic focusc</td>
<td>12.7</td>
<td>328.6</td>
</tr>
<tr>
<td>Mesoendemic focusd</td>
<td>3.5</td>
<td>369.7</td>
</tr>
<tr>
<td>Hypoendemic focusd</td>
<td>0.4</td>
<td>37.1</td>
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</table>

Table II. Results of the ELISA and ICT test for the detection of circulating antigen of Wuchereria bancrofti, according to three endemic levels in Pacific islands.
sified as good, at the inferior limit (κ = 0.60). In comparison with the ELISA reference test, the sensitivity and specificity of the ICT test were 51.2 % and 98.4 %, respectively. The results of ICT testing in the three endemic situations (hyperendemic in Tahaa, mesoendemic in Opoa and hypoendemic in Maupiti) are shown in Table II. A non negligible proportion of false negative results was noted, especially in situation with a high prevalence of microfilaraemia, adult worm carriers (14.5 % of false negative results in the Opoa cohort, where 26 % of the sample is made of such adult worm carriers). It is interesting to notice that the mean level of antigenemia in these false negative (Og4C3*, ICT) sera (533.7 ± 402.8 UI/mL, n = 32) was significantly (t = 6.34, p < 0.0001) lower than the one observed in the sample of Og4C3* sera (1,082.5 ± 603.2 UI/mL, n = 203). Consequently the degree of agreement with the ELISA data was the lowest (72.5 %) in the Opoa cohort, by comparison with the Tahaa and the Maupiti cohorts (90.0 % and 95.3 % of agreement, respectively). Similarly, the sensitivity and specificity were respectively 64.8 % and 97.9 % in Tahaa; 46.3 % and 99.3 % in Opoa; and 21.9 % and 98.4 % in Maupiti. Although the specificity of ICT test remained very good in all endemic situation, its sensitivity decreased (although not significantly) when the microfilarial prevalence decreased. According to the parasitological data, the degree of agreement with ELISA was good in both the microfilaremic and the amicrofilaremic group (data not shown). On the sub-sample of 50 reference sera, the agreement observed between the first-generation (serum-based) and the second-generation (whole blood-based) card tests was perfect (100 %). No rheumatoid factor-positive sera were positive with the ICT test.

On a practical basis, the card-test was very simple and straightforward to perform and seemed appropriate for a non-specialist laboratory, even in field conditions where small numbers of sera are generally to be tested every day. According to the manufacturer’s recommendations it seems better to store kits at + 2 to + 8 °C, even if the shelf life at + 25 °C is at least of six months, so refrigeration is recommended even if not absolutely necessary.

**DISCUSSION**

In order to evaluate the reliability of new rapid tests under local conditions, including different levels of parasitemia, we tested stored sera from three carefully managed human cohorts using the card-test and the standard ELISA assay. If the agreement between the two ICT tests (serum based- and whole blood based-tests) was good, their decreased sensitivity by comparison with the ELISA-based reference test was impai-

ring their use for individual diagnosis. In particular, it seems that the sensitivity of the ELISA (67 % in a brazillian focus, see Rocha et al, 1996) and the ICT (about 56 % in our polynesian focus) tests for detecting adult worm carriers is not perfect. Consequently, ICT tests can be better used to monitor at the population level the efficiency of filariasis control strategy, since we know that antigenemia decreased when control strategy is correctly conducted, according to our monitoring experiences of some pilot areas in French Polynesia (Moulia-Pelat et al., 1995; Nicolas, 1997).

Regarding the filariasis control programs recommended in different endemic areas by WHO, it is matter of great importance to assess their impact (WHO, 1996, 1998; Ottesen et al., 1997). Because laboratory facilities are not always available in endemic areas, and, moreover, because the standard techniques (ELISA, even when using a filter-paper version (Simonsen & Dunyo, 1999) or membrane filtration technique) are expensive and time-consuming, the use of ICT kits appears as a rapid and adequate tool, easy to perform, for the determination of filarial infection prevalence (Weil et al., 1997). This technique allows blood or plasma analysis in less than 15 minutes, and they can be taken day or night, which is logistically preferable. However, when testing sera from a focus with controlled transmission (i.e. with a low level of microfilaremia associated to high level of antigenemia, including a non negligible proportion of amicrofilaremic, adult worm carriers), there was an increasing risk of false negative results and the test was considered less useful in this peculiar situation. This could lead to a problem for stopping control programmes, as appropriate endpoints could be more difficult to define than previously thought.

**ACKNOWLEDGEMENTS**

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**REFERENCES**


